Acute Toxicity of Ammonia and Nitrite to Sea Bream, *Sparus aurata* (Linnaeus, 1758), in Relation to Salinity

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Abstract

Sea bream, *Sparus aurata*, is one of the most important fish species that is commonly cultured in the Mediterranean and the eastern coasts of the Atlantic Ocean. The life cycle of sea bream in its natural habitat passes through hyposaline and hypersaline lagoons. It is important to determine the tolerance of the fish to nitrogenous compounds for aquaculture at maximum stocking densities. In the present study, a series of acute experiments were performed to evaluate the effect of salinity on ammonia and nitrite toxicity to sea bream. The fish were exposed to different ammonia and nitrite concentrations according to the static renewal methodology at three different salinities (10, 20, and 30 ppt) and at a temperature of 20 C and a pH of 8.2. The toxic effect of total ammonia nitrogen (TAN) and nitrite nitrogen (NO₂-N) decreased with increasing salinity levels (P < 0.001). Acute toxicity (96-h lethal concentration 50 [LC₅₀]) values of TAN were determined to be 5.93, 11.72, and 19.38 mg/L at 10, 20, and 30 ppt salinity, respectively. The 96-h LC₅₀ values of NO₂-N were determined to be 370.80, 619.47, and 806.33 mg/L at 10, 20, and 30 ppt salinity, respectively. Results indicate that sea bream is less tolerant to ammonia but more tolerant to nitrite compared with some other fish species.

KEYWORDS

ammonia, LC₅₀, nitrite, salinity, Sparus aurata

Ammonia and nitrite can concentrate in aquaculture systems and cause commonly known water-quality problems (Frias-Espericueta et al. 1999). However, high densities of ammonia and nitrite concentration may increase to levels that reduce fish growth or even cause death. The toxic effect of ammonia on aquatic animals is ascribed to the unionized form of total ammonia nitrogen (TAN) (NH₃-N). The ionized form (NH_4^+-N) is considered nontoxic because it cannot pass through cell membranes. The NH₃-N that penetrates cell membrane destroys cell functions and causes death in fish (Jeney et al. 1992). Nitrite spreads into red blood cells and oxidizes hemoglobin to methemoglobin, which does not carry oxygen in blood, causing fish to die from anoxia (Lewis and Morris 1986; Jensen 1995).

Regarding the toxic effect of nitrogenous compounds in marine fish, salinity is one of the most important environmental variables that must be considered. High salinity levels increase tolerance of most aquatic organisms to ammonia and nitrite (Saroglia et al. 1981; Kır and Öz 2015). It has been reported that the toxic effect of nitrite on fish constantly decreases with increasing salinity (Saroglia et al. 1981; Wise and Tomasso 1989). Ammonia toxicity also decreases with increasing salinity up to levels of 30–40 ppt but then increases at higher salinity levels (Herbert and Shurben 1965; Harader and Allen 1983).

Sea bream, *Sparus aurata*, is a euryhaline fish species that is widely distributed in hyposaline and hypersaline lagoons around the Mediterranean and the eastern Atlantic Ocean (Polo et al. 1991; Vargas-Chacoff et al. 2009; Georgakopoulou et al. 2010). Various salinity levels are necessary in sea bream reproductive life cycles to keep larvae alive and to maintain homoeostasis. Gonadal growth and maturation of sea bream take place in estuaries and coastal waters (Arias 1976). In the winter season, mature sea bream migrate from lagoons and

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coastal waters to open seas for spawning where salinity conditions are unknown (Barbaro and Francescon 1986).

The culture of sea bream from the larval stage to juvenile size is mainly carried out in recirculating aquaculture systems. The accumulation of ammonia and nitrite needs to be considered in recirculating aquaculture systems along with salinity influences on the toxic effect of the nitrogenous compounds. It is important to measure the tolerance of fish to nitrogenous substances related to salinity. Although there have been some studies about the toxicity of ammonia and nitrite on sea bream related to temperature and pH (Wajsbrot et al. 1991, 1993; Parra and Yufera 1999), there is not much information on the harmful effects of nitrogenous compounds on sea bream at different salinity levels. The objective of this study was to determine the toxic effects of ammonia and nitrite on sea bream in relation to salinity. The present study estimated the median lethal time (LT_{50}) and the median lethal concentration (LC_{50}) of ammonia and nitrite for sea bream at different salinity levels in acute toxicity experiments.

Materials and Methods

Experimental Fish and Adaptation Procedure

A total of 720 $(1.1 \pm 0.2 \text{ g})$ juvenile sea bream were obtained for this study from the fish farm of Kılıç Holding near Bafa Lake in Turkey. Fish were transferred in water at a temperature of 20 C and 30 ppt salinity. Experiments were performed in the laboratory of the Aquaculture Department of the Faculty of Fisheries at Mugla Sitki Kocman University. After transfer to the laboratory, ambient salinity levels were reduced from 30 to 20 ppt at a rate of 5 ppt per day by adding fresh water. Fish were stocked in a 250-L tank that was part of a recirculating system at a stocking density of 1 kg/ton, and fish were maintained for 1 wk in this condition. Fish were then divided randomly into three groups, and each group was assigned to a 100-L fiberglass tank at 20 C. The salinity in the three tanks was increased or decreased at a rate of 2 ppt per day by adding fresh water or salt water until the desired final salinity levels (10, 20, and 30 ppt) were reached. Each group of fish was maintained at these different salinity levels for 2 wk and fed *ad libitum* with commercial feed pellets containing 45% protein. Acute ammonia and nitrite toxicity experiments were then performed to determine the LT_{50} and LC_{50} of the fish at different exposure times (24, 48, 72, and 96 h).

Toxicity Experiments

Ammonia and nitrite stock solutions (30 g/L) were prepared with ammonium chloride (NH₄Cl; Merck, Darmstadt, Germany) and sodium nitrite (NaNO₂; Merck) according to Chen and Lei (1990). The stock solutions were then diluted to the predicted TAN and NO₂-N concentrations with 10, 20, and 30 ppt seawater. The experimental TAN concentrations in the test solutions were 6, 8, 10, 12, 14, 16, 20, and 24 mg/L. Nominal concentrations of NO₂-N were 600, 700, 800, 900, 1000, 1100, and 1200 mg/L. The actual concentration levels of TAN and NO₂-N in test solutions were checked using methods reported by Parsons et al. (1985) and Bendschneider and Robinson (1952), respectively. The NH₃-N concentration in the TAN was calculated as described by Bower and Bidwell (1978) based on a pH of 8.2 with salinities of 10, 20, and 30 ppt at a temperature of 20 C. The salinity and pH of the test solutions were regularly controlled using an HQ40d multiparameter meter (Hach, Loveland, CO, USA).

The acute toxicity experiments were performed as described by APHA (1989). Ten fish were randomly sampled from each salinity group and were allocated to polyethylene test aquariums. Each aquarium contained 5L of test solution. Bioassay experiments were conducted in two replicates. Each test aquarium was aerated continuously. The test solution in each test aquarium was changed daily according to the static renewal methodology (Buikema et al. 1982). The fish were starved for 8 h before experimental trials and not fed during the experiment (96 h). The test fish were observed at 12-h intervals until the end of the trial. Fish that were motionless and without opercular movement were presumed dead. The LT_{50} and LC_{50} values

				1	ΓAN			NO ₂ -N				
	0.95	$df\chi^2$	п	χ^2	R^2	Regression equation	0.95	$df\chi^2$	п	χ^2	R^2	Regression equation
10 pp	t											
24 h	7.815	5	3	0.998	0.989	y = 3.16x + 1.27	3.841	1	3	0.106	0.984	y = 2.04x - 0.02
48 h	7.815	5	3	0.905	0.937	y = 4.60x + 0.07	3.841	1	3	0.775	0.994	y = 1.98x - 0.01
72 h	5.991	5	3	0.928	0.931	y = 3.03x + 2.18	3.841	1	3	0.662	0.935	y = 2.07x - 0.01
96 h	5.991	5	3	0.992	0.947	y = 3.40x + 2.35	3.841	1	3	0.787	0.960	y = 2.22x - 0.01
20 pp	t					·						-
24 h	5.991	4	2	0.003	0.589	y = 2.55x + 1.13	3.841	1	3	0.367	0.968	y = 12.7x - 32.7
48 h	5.991	4	2	0.309	0.959	y = 3.45x + 0.56	5.991	2	4	0.698	0.875	y = 11.9x - 29.2
72 h	5.991	5	3	0.668	0.962	y = 3.82x + 0.47	3.841	1	3	0.662	0.935	y = 11.0x - 26.1
96 h	5.991	5	3	0.834	0.893	y = 3.81x + 0.93	3.841	1	3	0.787	0.960	y = 12.3x - 29.5
30 pp	t											
24 h	5.991	5	3	0.632	0.929	y = 2.27x + 1.48	5.991	2	4	0.739	0.976	y = 9.96x - 24.8
48 h	5.991	5	3	0.592	0.992	y = 2.43x + 1.54	5.991	2	4	0.842	0.985	y = 7.50x - 17.1
72 h	5.991	5	3	0.537	0.908	y = 2.73x + 1.30	5.991	2	4	0.893	0.940	y = 7.85x - 17.9
96 h	5.991	5	3	0.507	0.956	y = 2.95x + 1.20	5.991	2	4	0.937	0.952	y = 8.34x - 19.2

TABLE 1. The relationship between probit mortality (y) and the log of NO_2 -N and total ammonia nitrogen (TAN) concentrations in mg/L (x) at various exposure times and salinity levels for Sparus aurata.

of ammonia and nitrite concentrations were calculated according to the Bliss Probit Method. The 95% confidence limits of the LT_{50} and LC_{50} values were estimated (Sprague 1969). The probit mortality line, which is a function of the log of TAN and NO₂-N concentrations, was determined, and a chi-square (χ^2) test was performed to determine the goodness of fit.

Statistical Analysis

The SPSS 22 statistical software (IBM, New York, NY) was used for two-way ANOVA testing to compare differences between groups based on salinity, survival, concentrations of toxic substances, and exposure time. Duncan's multiple range tests were performed to compare the mean values of LC_{50} among salinities and exposure times. Regression analyses were used to determine relationships of TAN and NO₂-N concentrations and exposure time on LT_{50} and LC_{50} values.

Results

Throughout the ammonia and nitrite toxicity experiments, no deaths occurred in the control solutions at any salinity levels. Low levels of nitrogen compounds were not toxic to the fish, whereas high concentrations had a lethal effect at even 12 h of exposure. The estimated probit mortalities are illustrated in Table 1. It was determined from the results of the χ^2 analysis that there is a positive linear relationship between the log of ammonia and nitrite concentrations and probit mortality. The LC₅₀ values of the toxic substances used in the study and their calculated 95% confidence limits for juvenile sea bream at different salinity levels and exposure times are summarized in Table 2. The LC₅₀ values for TAN and NO₂-N were 2.7 and 1.7 times higher, on average, at 30 ppt salinity than at 10 ppt salinity, respectively, at 24, 48, 72, and 96 h of exposure time.

The LT_{50} values estimated at different TAN and NO₂-N concentrations in the toxicity experiments are illustrated in Figure 1. The regression analyses performed in the study demonstrated a negative exponential relationship between the LT_{50} value and TAN and NO₂-N concentrations. The LT_{50} values varied depending on concentrations of TAN from 75.3 to 15.39 h and 175.3 to 36.02 h at 8–16 and 12–20 mg/L and 10 and 30 ppt salinity, respectively. The LT_{50} values varied depending on concentrations of NO₂-N from 47.5 to 5.7 h and 116.1 to 6.05 h at 600–800 and 800–1200 mg/L and 10 and 30 ppt salinity, respectively.

Discussion

The results of the present study clearly demonstrated that the tolerance of fish to toxic

Salinity (ppt)	TAN LC ₅₀	NH ₃ -N LC ₅₀	NO ₂ -N LC ₅₀	
10 ppt				
24 h	15.36 (14.0-16.7)	0.65 (0.60-0.71)	674.42 (673.3-675.5)	
48 h	11.66 (10.4–12.9)	0.50 (0.44-0.55)	573.34 (572.2-574.5)	
72 h	8.49 (7.13-9.84)	0.36 (0.30-0.42)	496.26 (496.3-497.4)	
96 h	5.93 (4.54-7.32)	0.25 (0.19-0.31)	370.80 (369.5-372.1)	
20 ppt				
24 h	33.39 (31.9-34.8)	1.42 (1.36-1.48)	921.26 (920.2-922.3)	
48 h	19.23 (17.9–20.5)	0.82 (0.76-0.87)	748.92 (747.8-750.0)	
72 h	15.21 (14.0-16.5)	0.65 (0.59-0.70)	656.07 (655.0-657.2)	
96 h	11.72 (10.5-13.0)	0.50 (0.45-0.55)	619.47 (618.4-620.6)	
30 ppt				
24 h	42.68 (41.0-44.3)	1.66 (1.60-1.73)	990.27 (989.2-991.4)	
48 h	26.32 (24.9-27.7)	1.03 (0.97-1.08)	887.16 (886.0-888.3)	
72 h	22.52 (21.2-23.9)	0.88 (0.82-0.93)	836.02 (834.9-837.1)	
96 h	19.38 (18.0-20.7)	0.76 (0.70-0.81)	806.33 (805.2-807.4)	

TABLE 2. Lethal dose 50 (LC_{50}) (mg/L) of total ammonia nitrogen (TAN), NH₃, and NO₂-N for Sparus aurata and 95% confidence limits (in parenthesis) at three salinity levels and various exposure times.^{*a*}

^aThe LC₅₀ value in the same column and row are different from each other (P < 0.001).

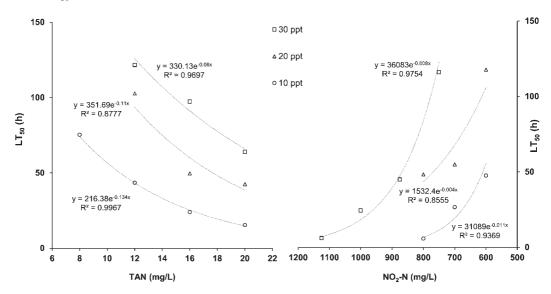


FIGURE 1. Lethal time (LT_{50}) (h) values for Sparus aurata exposed to different concentrations of total ammonia nitrogen (TAN) and NO₂-N at a salinity of 10, 20, and 30 ppt.

substances increases as salinity levels increase (P < 0.001). Similar results related to salinity have also been reported in studies on the European eel, *Anquilla anquilla* (Saroglia et al. 1981) and the grey mullet, *Mugil planatus* (Sampaio et al. 2002).

There are some previous studies on the toxic effects of ammonia on sea bream at various salinity levels and at various temperatures with respect to size or developmental stage. For example, a 57-mg/L 96-h LC₅₀ value of TAN was reported for sea bream at 6-160 g in weight at 34 ppt salinity, 18 C, and a pH of 8.1 (Le Ruyet et al. 1995). For 12-d-old larvae of sea bream, the 24-h LC₅₀ values of NH₃-N was reported as 0.24 mg/L at 33 ppt salinity, 19.5 C, and a pH of 8.2 (Parra and Yufera 1999). While the 96-h LC₅₀ value for sea bream of 1.1 g was calculated as 19.38 mg/L for TAN, the 24-h LC₅₀ value of NH₃-N was calculated as 1.66 mg/L at 30 ppt

salinity, 20 C, and a pH of 8.2 in the current study. The results of the studies carried out previously on sea bream and the results obtained from this study demonstrate that large fish have a high tolerance to ammonia compared to small fish.

The 96-h LC₅₀ value of meagre, Argyrosomus regius, with a weight of 3.0 g, has been reported as 10.39 mg/L TAN at 30 ppt salinity, 22 C, and a pH of 8.2 (Kir et al. 2016). European sea bass, Dicentrarchus labrax, and turbot, Scopthalmus maximus, with weights ranging from 6 to 160 g, have 96-h LC₅₀ values of 40 and 59 mg/L TAN, respectively, at similar salinity, temperature, and pH (34 ppt, 18 C, and 8.1) (Le Ruyet et al. 1995). At a salinity of 30 ppt, temperature of 20 C, and pH of 8.2, the 96-h LC₅₀ value of TAN for sea bream (at 1.1 g) was calculated as 19.38 mg/L in the current study. Temperature, salinity, and the size or development stage of fish may affect the response of fish to ammonia (Kır et al. 2016). In addition, it has been suggested that ammonia may interact with other environmental variables, such as nitrite, oxygen, and carbon dioxide (Randall and Wright 1987; Wajsbrot et al. 1991; Foss et al. 2003; Lemarie et al. 2004). The results of previous studies on other fish species and the results of the current study indicate that sea bream is a fish that is less tolerant to ammonia than some other fish species cultured in the Mediterranean and eastern Atlantic, such as European sea bass and turbot, but sea bream is more tolerant than meagre.

The 96-h LC₅₀ values of NO₂-N for juvenile sea bream were calculated as 370, 619, and 806 mg/L at 10, 20, and 30 ppt salinity, respectively, at a temperature of 20 C and a pH of 8.2 in the current study (Table 2). Kir et al. (2015) reported a 96-h LC_{50} value of NO_2 -N for 5.4-cm meagre as 139.5 mg/L at 22 C and 30 ppt salinity. For European sea bass, with a length of 5 cm, a 96-h LC_{50} value was reported as 220 mg/L NO₂-N at 23 C. In the same study, a 96-h LC_{50} value for European eel, with a length of 7 cm, was reported as 812 mg/L NO2-N at 30 ppt salinity (Saroglia et al. 1981). Such significant differences in the 96-h LC₅₀ value of NO₂-N among fish species indicate that each fish has species-specific regulatory mechanisms for nitrite (Wise and Tomasso 1989). In all conditions, the present study demonstrated that sea bream has nitrite sensitivity that is similar to the European eel, but sea bream is more tolerant than European sea bass and meagre.

The 24-h LC₅₀ value of NO₂-N for 12-d-old sea bream larvae has been reported as 1997 mg/L at 33 ppt salinity, a temperature of 19.5 C, and a pH of 8.2 (Parra and Yufera 1999). In the current study, the 24-h LC₅₀ value of NO₂-N was calculated as 990 mg/L for juvenile sea bream at salinity of 30 ppt, temperature of 20 C, and pH of 8.2. Studies have shown that small fish and larvae are less tolerant to nitrite compared with large fish of the same species (Lewis and Morris 1986). However, the LC_{50} value estimated in the previous study for sea bream larvae is higher than the LC₅₀ value calculated in the current study for juvenile sea bream at 1.1 g. In both studies, toxicity experiments were performed at similar temperatures and pH. Because there were salinity differences, it is thought the LC_{50} value of larvae is higher than the LC_{50} value of juveniles.

A safe level is defined as an amount of toxic substance that has no adverse effect on aquatic organisms, and it is calculated by dividing a 96-h LC₅₀ by a factor of 10 (Sprague 1971). Safe levels are generally considered safe concentrations in which organisms can remain alive and grow. The safe levels of TAN for sea bream were estimated to be 0.5, 1.1, and 1.9 mg/L at 10, 20, and 30 ppt salinity, respectively. The NO₂-N safe levels were estimated to be 37, 61, and 80 mg/L at 10, 20, and 30 ppt salinity, respectively (P < 0.001). From the current data, the toxic effects of TAN and NO₂-N on sea bream appear to be 4.5 and 2.2 times higher, respectively, at a salinity of 10 ppt compared with 30 ppt. The safe levels estimated in this study are important factors to be considered for better management of sea bream culture, particularly when considering different salinities. It is necessary to prevent the accumulation of these nitrogenous compounds at concentrations higher than safe levels in production systems.

Conclusions

The toxic effect of ammonia and nitrite on sea bream juveniles decreases with increasing

salinity. When a comparison is made among fish species cultured in the Mediterranean and eastern Atlantic coasts, sea bream is a less tolerant fish species to ammonia compared with European sea bass, but it is more tolerant than meagre. Regarding nitrite toxicity, sea bream has more tolerance compared with the other two fish species.

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