

# COMPARISON OF ANTIBIOTIC SENSITIVITY RATIO OF *ESHERICHIA COLI* AND *KLEBSIELLA PNEUMONIAE* STRAINS ISOLATED FROM VARIOUS CLINICAL SPECIMENS IN INTENSIVE CARE UNIT

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## ABSTRACT

A retrospective analysis of the widely used antibiotics all susceptibility testing results from *Escherichia coli* and *Klebsiella pneumoniae* cultured from clinical specimens Muğla Sitki Koçman University Education and Research hospital (January to December in 2015) was performed. The new BD PHOENIX automated microbiology system (Becton Dickinson Diagnostic Systems, Sparks, Md.) is designed for automated rapid antimicrobial susceptibility testing and identification of clinically relevant bacteria. Minimum Inhibitory Concentration (MIC) results previously obtained in recent clinical isolates with well-defined in isolates with well-characterized resistance mechanisms with the microdilution method were re-interpreted for the susceptible, intermediate and resistant categories using the 2012 EUCAST breakpoints. Clinical samples are most commonly isolated from tracheal aspirates, wound site, blood, mucus, abscess, catheter, urine samples, throat and nose cultures. *E.coli* isolates were highly resistant to Ciprofloxacin (CIP), Cefepime (FEP), and Aztreonam (ATM) 47%, 47% and 40% respectively. Unlike *K.pneumoniae* isolates were highly resistant to Ampicillin/Sulbactam (SAM), ATM and FEP 58%, 53% and 50% respectively. Both of the bacteria showed the highest sensitivity rate to Amikacin (AN) 98% and 90% *E.coli* and *K.pneumoniae* respectively. Out of the 60 *E.coli* strains 34 (57%) isolates showed Multiple Antibiotic Resistance (MAR) two to ten antibiotics. Out of the 60 *K.pneumoniae* strains 38 (63%) isolates showed MAR two to ten antibiotics. Considering the antibiogram, AN and Meropenem (MEM) should be preferred drugs for *K.pneumoniae* and *E.coli* infection isolated from clinical samples.

## KEYWORDS:

*Escherichia coli*, *Klebsiella pneumoniae*, antibiotic sensitivity, clinical specimens, intensive care unit

## INTRODUCTION

Antibiotic resistance in Gram-negative bacteria is a major threat to public health [1-4]. Patients with non-severe infections caused by multidrug-resistant bacteria are subject to in-hospital intravenous therapy because there are no effective oral drugs available. Resistance to empirical antibiotic therapy results in delayed appropriate antibiotic treatment for severe infections, which is associated with increased mortality, prolonged hospital stay and higher costs [5-7]. Further, resistance challenges the achievements of modern medicine, including advanced surgery and immunosuppressive treatment, which are dependent on effective antibiotics.

*Escherichia coli* and *Klebsiella pneumoniae* are two important members of Gram-negative rods (Enterobacteriaceae), which belongs to a part of human gastrointestinal normal flora [8-10]. These organisms are substantial human pathogens which lead to a wide spectrum of hospital and community-acquired infections such as urinary tract infection (UTI), septicemia, pneumonia, peritonitis, meningitis, etc. [11, 12]. In the lack of appropriate treatment of infections by these organisms, noticeable morbidity and mortalities will occur [13].

The aim of this study was to determine the characteristics and patterns of antibiotic resistance among isolates of *Escherichia coli* and *Klebsiella pneumoniae* recovered from clinical specimens in Muğla

## MATERIALS AND METHODS

**Bacterial isolates.** Ethical approval is was taken before study. Because of retrospective analysis, we did not patient approval. 60 *Escherichia coli* and 60 *Klebsiella pneumoniae* were isolated from clinical specimens from intensive care unit of internal medicine in Muğla Sitki Koçman University Education and Research hospital. Bacterial isolates were identified to level of species and subspecies by using the morphological and traditional biochemical

tests and automatic diagnostic systems currently present in the market and commonly used for AST (Antimicrobial Susceptibility Testing) in clinical laboratories will therefore have to incorporate these criteria in their instruments to meet the needs of European microbiology laboratories according to standard methods described by [14-16]. All isolates were obtained from patients at intensive care units. In total, 120 *Escherichia coli* and *Klebsiella pneumoniae* were isolated from various clinical samples and detected by the PHOENIX (Becton Dickinson, USA) at the microbiology laboratory of our hospital between from January to December 2015. The Phoenix™ Automated Microbiology System (BD Diagnostics, Sparks, MD, USA) is designed for the rapid bacterial identification at the species level and determination of AST of clinically significant human bacterial pathogens [17].

**Antibiogram profile of *Escherichia coli* and *Klebsiella pneumoniae*.** Minimum Inhibitory Concentration (MIC) results previously obtained in recent clinical isolates with well-defined in isolates with well-characterized resistance mechanisms with microdilution method were re-interpreted for the susceptible, intermediate and resistant categories using the 2012 EUCAST breakpoints. Ten different antibiotics were used.

**Multiple Antibiotic Resistance (MAR) index.** For all isolates, we calculated the MAR index values (a/b, where a represents the number of antibiotics the isolate was resistant to, b represents the total number of antibiotics the isolate tested against). A MAR index value  $\geq 0.2$  is observed when isolates are exposed to high risk sources of human or animal contamination, where antibiotics use is common; in contrast a MAR index value  $< 0.2$  observed when antibiotics are seldom or never used [18-19].

## RESULTS AND DISCUSSION

**Resistance pattern results.** The results of resistance pattern of *E.coli* isolates in our locality to antimicrobial agents showed that the 60 *E.coli* and 60 *K.pneumoniae* strains tested against ten antimicrobial agents in Table 1. *E.coli* isolates were highly resistant to Ciprofloxacin (CIP), Cefepime (FEP), and Aztreonam (ATM) 47%, 47 and 40% respectively. Unlike *K.pneumoniae* isolates were highly resistant to Ampicillin/Sulbactam (SAM), Aztreonam (ATM) and Cefepime (FEP) 58%, 53% and 50% respectively. Both of the bacteria showed the highest sensitivity rate to Amikacin (AN) 98% and 90% *E.coli* and *K.pneumoniae* respectively.

In our study, when we compared to resistance of CIP, *E.coli* isolates showed high antibiotic resistance with 47% CIP. Unlike *K.pneumoniae* isolates showed resistance to CIP with 18%. Many researchers were reported resistance rate to CIP *E.coli* and *K.pneumoniae* [20-22]. Our results were similar to Giray et al. (2012) [23] who also reported that Ciprofloxacin resistance rate to *E.coli* was 47%.

In this study, resistance of FEP *E.coli* and *K.pneumoniae* 47% and 50% respectively. Many researchers were reported resistance rate to FEP *E.coli* and *K.pneumoniae* [24-27]. Our results were similar to previous researchers.

When it comes to resistance of ATM *K.pneumoniae* isolates showed high resistance to ATM with 53%. Unlike *E.coli* isolates showed resistance to ATM with 40%. Many researchers were reported resistance rate to ATM *E.coli* and *K.pneumoniae* [25-33]. Our results were similar to Sabir et al. (2014) [33] who also reported that ATM resistance rate to *E.coli* was 44.8%.

In our study, resistance of Norfloxacin (NOR) were *E.coli* with 35% and *K.pneumoniae* with 32%. Many researchers were reported resistance rate to NOR *E.coli* and *K.pneumoniae* [34-36]. Our results were similar to previous researchers.

**TABLE 1**  
**Antibiotic resistance pattern of *E.coli* and *K.pneumoniae* isolated from clinical specimens in intensive care unit.**

Antibiotics	<i>E.coli</i>			<i>K.pneumoniae</i>		
	R	I	S	R	I	S
CIP	28(47%)	-	32(53%)	23(18%)	1(2%)	36(60%)
FEP	28(47%)	2(3%)	30(50%)	30(50%)	1(2%)	29(48%)
ATM	24(40%)	-	36(60%)	32(53%)	-	28(47%)
NOR	21(35%)	-	39(65%)	19(32%)	1(2%)	40(67%)
CXM	20(33%)	1(2%)	39(65%)	28(47%)	1(2%)	31(52%)
SAM	18(30%)	14(23%)	28(47%)	35(58%)	5(18%)	20(33%)
TZP	6(10%)	4(7%)	50(83%)	14(23%)	7(12%)	39(65%)
AMC	5(8%)	8(13%)	47(78%)	13(22%)	9(15%)	38(63%)
MEM	3(5%)	2(3%)	55(92%)	15(25%)	3(5%)	42(70%)
AN	1(2%)	-	59(98%)	6(10%)	-	54(90%)

Abbreviations; CIP; Ciprofloxacin, FEP; Cefepime, ATM; Aztreonam, NOR; Norfloxacin, CXM; Cefuroxime sodium, SAM; Ampicillin/Sulbactam; TZP; Tazobactam/Piperacillin, AMC; Amoxicillin/Clavulanic Acid, MEM; Meropenem, AN; Amikacin, -; No result.

**TABLE 2**  
**Number of clinical samples and MAR Index 60 *E.coli* and *K.pneumoniae* strains.**

Clinical Samples in Intensive Care Unit	<i>E.coli</i>				<i>K.pneumoniae</i>			
	Number of Isolates	MAR			Number of Isolates	MAR		
Urine	37	0(13isl), 0,1(4isl), 0,2(7isl), 0,3(1), 0,4(2isl), 0,5(3isl), 0,6(4isl), 0,7(1), 0,8(2isl)			35	0(9isl), 0,1(4isl), 0,2(3isl), 0,3 (3isl), 0,4(3isl), 0,5(1isl), 0,6 (5isl), 0,7(3isl), 0,8(1isl), 0,9(2isl), 1(1isl)		
Tracheal aspirate	6	0(1), 0,1(1isl), 0,5(2isl), 0,6(1isl), 0,7(1isl)			10	0(4isl), 0,2(2isl), 0,3(1isl), 0,4(1isl), 0,5(1isl), 0,8(1isl)		
Blood	4	0,3(2isl), 0,4(2isl)			5	0(2isl), 1(1isl), 0,5(1isl), 0,9(1isl)		
Gaita	1	0,4			-			
Vagen	6	0(2isl), 0,1(3isl), 0,2(1isl)			2	0(1isl), 0,5(1isl)		
Abces	4	0(1isl), 0,1(1isl); 0,2(1isl); 0,3(1isl)			2	0(1isl), 0,5(1isl)		
Wound	1	0,3(1isl)			3	0,1(1isl), 0,8(1isl), 1(1isl)		
Periton	1	0,3(1isl)			-			
Throat	-	-			1	0,4(1isl)		
Mucus	-	-			2	0,4(1isl), 0,7(1isl)		

MAR, Multiple Antibiotic Resistance Index, isl; isolates, -; No result.

When we compared to resistance of Cefuroxime sodium (CXM) *K.pneumoniae* isolates showed high resistance to CXM with 47%, but *E.coli* isolates showed resistance of CXM with 33%. Many researchers were reported resistance rate to CXM *E.coli* and *K.pneumoniae* [30, 37]. Our results were similar to previous researchers.

In this study, *K.pneumoniae* resistance (58%) of SAM were higher than *E.coli* (30%). Many researchers were reported resistance rate to SAM *E.coli* and *K.pneumoniae* [38-42]. Our results were similar to Tsakris et al. (1997) [42] who also reported that SAM resistance rate to *E.coli* was 32.1%.

When we compared to resistance of Tazobactam/Piperacillin (TZP), *K.pneumoniae* (23%) isolates showed more resistance than *E.coli* (10%). Many researchers were reported resistance rate to TZP *E.coli* and *K.pneumoniae* [43, 44]. Our results were similar to Tsakris et al. (1997) [42] who also reported that TZP resistance rate to *E.coli* was 10.5%.

When it comes to resistance of Amoxicillin/Clavulanic acid (AMC), *K.pneumoniae* (22%) isolates showed more resistance than *E.coli* isolates (8%). Many researchers were reported resistance rate to AMC *E.coli* and *K.pneumoniae* [45-47]. Our results were similar to previous researchers.

In this research, Meropenem (MEM) and AN were the most sensitive antibiotics to both of the isolates. While percentage of MEM sensitivity was 92% and 70% *E.coli* and *K.pneumoniae* respectively. Many researchers were reported resistance rate to MEM *E.coli* and *K.pneumoniae* [36, 48-50]. Our results were similar to Sohail et al. (2015) [51] who also reported that MEM resistance rate to *E.coli* was 3%.

In our study, sensitivity rate of AN were *E.coli* with 98% and *K.pneumoniae* with 90%. Many researchers were reported resistance rate to AN *E.coli* and *K.pneumoniae* [23,31,52-54]. Manikandan and

Amsath (2013) [55] reported that the highest susceptibility pattern of bacterial isolates from respiratory tract infection showed to AN like our study.

**MAR index results.** Out of the 60 *E.coli* strains 34 (57%) isolates showed MAR two to ten antibiotics. 17 (28%) isolates showed no antibiotic resistance especially isolated from urine samples. Out of the 60 *K.pneumoniae* strains 38 (63%) isolates showed MAR two to ten antibiotics. Both of the bacteria, 17 (28%) isolates showed no antibiotic resistance especially isolated from urine samples. The MAR indices give an indirect suggestion of the probable source(s) of the organism. The results were given Table 2. Some researchers have reported MAR rate to *E.coli* from 2% to 97% [56-58]. MAR is considered as a good tool for risk assessment. This also gives an idea of the number of bacteria showing antibiotic resistance in the risk zone in the study's routine susceptibility testing. This MAR index also recommended that all isolates, somehow, originated from the environment where antibiotics were over used% [59]. Notably, MAR indices for the clinical isolates of *E. coli* were generally lower than *Klebsiella pneumoniae* isolates recovered from same conditions (Table 2).

## CONCLUSIONS

High antibiotic resistance of *K. pneumoniae* and *E.coli* towards commonly used antibiotics are the major reasons for prolonged infections, increased hospitalization, increased cost of therapy and enhanced morbidity and mortality rates. *K. pneumoniae* and *E.coli* was found to be most sensitive to AN and MEM. Considering the antibiogram, AN and MEM should be preferred drugs for *K. pneumoniae* and *E.coli* infection isolated from clinical samples. MAR indices for the clinical isolates of *E.*

*coli* were generally lower than *K. pneumoniae* isolates recovered from same conditions. The findings in present the study suggest that there is an urgent need for constant monitoring of susceptibility of pathogens in different populations to commonly used antimicrobial agents. The data of this study may be used to determine trends in antimicrobial susceptibilities, to formulate local antibiotic policies and overall to assist clinicians in the rational choice of antibiotic therapy to prevent misuse, or overuse, of antibiotics.

## REFERENCES

- [1] Kraker, M.E., Wolkewitz, M., Davey, P.G., Koller, W., Berger, J., Nagler, J., Icket, C., Kalenic, S., Horvatic, J., Seifert, H., Paniara, O., Argyropoulou, A. (2011) Burden of antimicrobial resistance in European hospitals: excess mortality and length of hospital stay associated with bloodstream infections due to *Escherichia coli* resistant to third-generation cephalosporins. *Journal of Antimicrobial Chemotherapy*. 66, 398-407.
- [2] Miyakis, S., Pefanis, A., Tsakris, A. (2011) The challenges of antimicrobial drug resistance in Greece. *Clinical Infectious Diseases*. 53, 177-184.
- [3] Ceyhan-Güvensen, N., Keskin, D., Sankur, F. (2017) Antibiotic resistance ratio of *Acinetobacter baumannii* against to ten antibiotics and multidrug resistance index in clinical isolates. *Fresen. Environ. Bull.* 26, 8316-8320.
- [4] Rossi, F. (2011) The challenges of antimicrobial resistance in Brazil. *Clinical Infectious Diseases*. 52, 1138-1143.
- [5] Schwaber, M.J., Carmeli, Y. (2007) Mortality and delay in effective therapy associated with extended-spectrum beta-lactamase production in Enterobacteriaceae bacteraemia: a systematic review and meta-analysis. *Journal of Antimicrobial Chemotherapy*. 60, 913-920.
- [6] Schwaber, M.J., Navon-Venezia, S., Kaye, K.S., Ben-Ami, R., Schwartz, D., Carmeli, Y. (2006) Clinical and economic impact of bacteraemia with extended- spectrum-beta-lactamase-producing Enterobacteriaceae. *Antimicrob Agents Chemother.* 50, 1257-1262.
- [7] Alharbi, N.S., Khaled, J.M., Kadaikunnan, S., Mothana, R., Alobaidi, A.S., Salmen, S. (2016) Evaluation of an efficiency of alcoholic extracts isolated from *Lagenaria siceraria* (Molina) against some antibiotic resistant clinical microorganisms. *Fresen. Environ. Bull.* 25, 3251-3256.
- [8] Paterson, D.L., Ko, W.C., Von Gottberg, A., Mohapatra, S., Casellas, J.M., Goossens, H., Mulazimoglu, L., Trenholme, G. (2004) Antibiotic therapy for *Klebsiella pneumoniae* bacteraemia: implications of production of extended-spectrum betalactamases. *Clinical Infectious Diseases*. 39, 31-37.
- [9] Ayatollahi, J., Vahidi, A., Shahcheraghi, S.H., Bagheripour, A., Lotfi, M., Lotfi, M., Lotfi, S.R. (2013) Investigating the resistance of *Escherichia coli* against some selected antimicrobials in Bam. *Jundishapur Journal of Microbiology*. 6(8), 7407-7411.
- [10] Dormanesh, B., Dehkordi, S.F., Hosseini, S., Momtaz, H., Mirnejad, R., Hoseini, M.J., Yahaghi, E., Tarhriz, V., Khodaverdi Darian, E. (2014) Virulence factors and o-serogroups profiles of uropathogenic *Escherichia coli* isolated from Iranian pediatric patients. *Iran Red Crescent Med J.* 16(2), 125-132.
- [11] Atmani, S.M., Messai, Y., Alouache, S. (2015) Virulence characteristics and genetic background of ESBL-producing *Klebsiella pneumoniae* isolates from wastewater. *Fresen. Environ. Bull.* 24, 103-112.
- [12] Allocati, N., Masulli, M., Alexeyev, M.F., Di Ilio, C. (2013) *Escherichia coli* in Europe: an overview. *International Journal of Environmental Research Public Health*. 10(12), 6235-54.
- [13] Russo, T. (2003) Medical and economic impact of extraintestinal infections due to *Escherichia coli*: focus on an increasingly important endemic problem. *Microbes Infectious*. 5(5), 449-56.
- [14] Cappuccino, J.G., Sherman, N. (2004) *Microbiology: A Laboratory Manual*. 7th Edn. Pearson Education (Singapore). Indian Branch, New Delhi, 544.
- [15] Gunasekaran, P. (2000) *Laboratory Manual in Microbiology*. New Age. International, New Delhi, 320.
- [16] MacFaddin, F.J. (2000) *Biochemical tests for identification of medical bacteria*. 3rd Edn. Philadelphia, USA, 912p.
- [17] O'Hara, C.M. (2005) *Manual and automated instrumentation for identification of Enterobacteriaceae and other aerobic gram-negative bacilli*. *Clinical Microbiology Reviews*. 18, 147-162.
- [18] Krumperman, P.H. (1985) Multiple antibiotic resistance indexing of *Escherichia coli* to identify high-risk sources of fecal contamination of foods. *Applied and Environmental Microbiology*. 46, 165-170.
- [19] Matyar, F., Kaya, A., Dinçer, S. (2008) Antibacterial agents and heavy metal resistance in Gram-negative bacteria isolated from seawater, shrimp and sediment in Iskenderun Bay, Turkey. *Science Total and Environment*. 407, 279-285.



- [20] Aiyegoro, O.A., Igbinsosa, O.O., Ogunmwonyi, I.N., Odjadre, E.E., Igbinsosa, O.E., Okoh, A.I. (2007) Incidence of urinary tract infections (UTI) among children and adolescents in Ile-Ife, Nigeria. *African J. Microbiol. Res.* 12, 013–019.
- [21] Turnidge, J., Bell, J., Biedenbach, J., Jones, R.N. (2002) Pathogen occurrence and antimicrobial resistance trends among urinary tract infection isolates in the Asia-Western Pacific Region: report from the SENTRY Antimicrobial Surveillance Program, 1998–1999. *International Journal of Antimicrobial Agents.* 20, 10–17.
- [22] Mansouri, S., Shareif, S., Ahmad, K. (2002) Antimicrobial resistance pattern of *E. coli* causing urinary tract infections and that of human faecal flora in southeast of Iran. *Iran Medical Journal.* 8, 123–128.
- [23] Giray, B., Ucar, F.B., Aydemir, S.S. (2012) Characterization of uropathogenic *Escherichia coli* strains obtained from urology outpatient clinic of Ege Medical Faculty in Izmir. *Turk J Med Sci.* 42(1), 1328–1337.
- [24] Kumarasinghe, G., Chow, C., Tambyah, P.A. (2001) Widespread resistance to new antimicrobials in a university hospital before clinical use. *International Journal of Antimicrob Agents.* 18(4), 391–393.
- [25] Iqbal, M., Patel, I.K., Shah, S.H., Ain, Q., Barney, N., Kiani, Q., Rabhani, K.Z., Zaidi, G., Mehdi, B. (2002) Susceptibility patterns of *Escherichia coli*: prevalence of multidrug-resistant isolates and extended spectrum beta-lactamase phenotype. *Journal of Pakistan Medical Association.* 52(9), 407–411.
- [26] Ashgar, A.H. (2006) Frequency and anti-microbial susceptibility patterns of bacterial pathogens isolated from septicemic patients in Makah hospitals. *Saudi Medical Journal.* 27(3), 361–367.
- [27] Albayrak, N., Kaya, S. (2009) Extended spectrum beta lactamases production and antimicrobial resistance ratio of the *Escherichia coli* and *Klebsiella pneumoniae* strains isolated from various clinical specimens. *Turkish Society of Microbiology Journal.* 39, 16–21.
- [28] Gonlugur, U., Bakici, M.Z., Akkurt, I., Efeoglu, T. (2004) Antibiotic susceptibility patterns among respiratory isolates of Gram negative bacilli in a Turkey university hospital. *BMC Microbiol.* 4, 1–5.
- [29] Cho, S.H., Lim, Y.S., Park, M.S., Kim, S.H., Yeon-Ho Kang, Y.H. (2011) Prevalence of antibiotic resistance in *Escherichia coli* fecal isolates from healthy persons and patients with diarrhea. *Public Health Res Perspect.* 2(1), 41–45.
- [30] Rashid, M., Rakib, M.M., Hasan, B. (2015) Antimicrobial-resistant and ESBL producing *Escherichia coli* in different ecological niches in Bangladesh. *Infection Ecology and Epidemiology.* 5, 26712.
- [31] Shilpa, K., Thomas, R., Ramyashree, A. (2016) Isolation and Antimicrobial sensitivity pattern of *Klebsiella pneumoniae* from sputum samples in a tertiary care hospital. *International Journal of Biomedical and Advance Research.* 7(2), 053–057.
- [32] Ruh, E., Gazi, U., Guvenir, M., Suer, K., Cakir, N. (2016) Antibiotic resistance rates of *Pseudomonas aeruginosa*, *Acinetobacter baumannii* and *Klebsiella pneumoniae* isolated from a university-affiliated hospital in North Cyprus. *Turk Hij Den Biyol Derg.* 73(4), 333–344.
- [33] Sabir, S., Anjum, A.A., Ijaz, T., Ali, M.A., Khan, M.R., Nawaz, M. (2014) Isolation and antibiotic susceptibility of *E. coli* from urinary tract infections in a tertiary care hospital. *Pakistan Journal of Medical Science.* 30(2), 389–392.
- [34] Karki, T., Truuaslu, K., Mikellsaar, M. (2001) Antibiotic susceptibility patterns of community- and hospital-acquired *Staphylococcus aureus* and *Escherichia coli* in Estonia. *Scand J Infect Dis.* 33, 333–338.
- [35] Ay, S., İşeri, L.A., Duman, B. (2003) Antibiotic susceptibilities of gram negative microorganisms isolated from urine samples. *İnönü Üniv Tıp Fak Derg.* 10(2), 59–62.
- [36] Amin, A., Ghumro, P.B., Hussain, S., Hameed, A. (2009) Prevalence of antibiotic resistance among clinical isolates of *Klebsiella pneumoniae* isolated from a Tertiary Care Hospital in Pakistan. *Malaysian Journal of Microbiology.* 5(2), 81–86.
- [37] Hatzaki, D., Poulakou, G., Katsarolis, I., Lambri, N., Souli, M., Deliolanis, I., Nikolo-poulos, G.K., Lebessi, E., Giamarellou, H. (2012) Cefditoren: Comparative efficacy with other antimicrobials and risk factors for resistance in clinical isolates causing UTIs in outpatients. *BMC Infectious Diseases.* 12, 228–234
- [38] Davis, G.S., Waits, K., Nordstrom, L., Weaver, B., Aziz, M. (2015) Intermingled *Klebsiella pneumoniae* populations between retail meats and human urinary tract infections. *Clinical Infectious Diseases.* 61(6), 892–899.
- [39] Toroglu, H., Avan, D. Keskin. (2013) Beta-Lactamases production and antimicrobial resistance ratio of *Pseudomonas aeruginosa* from hospitalized patients in Kahramanmaraş, Turkey. *Journal of Environmental Biology.* 34(4), 695–700.

- [40] Abuhandan, M., Güzel, B., Oymak, Y., Çiftçi, H. (2013) Antibiotic sensitivity and resistance in children with urinary tract infection in Sanliurfa. *Turkish Journal of Urology*. 39(2), 106-10.
- [41] Khan, A.U., Zaman, M.S. (2006) Multiple drug resistance pattern in urinary tract infection patients in Aligarh. *Biomedical Research*. 17(3), 179-181.
- [42] Tsakris, A., Douboyas, J., Leonidas, S. (1997) High rates of resistance to piperacillin/ tazobactam among *Escherichia coli* and *Klebsiella pneumoniae* strains isolated in a Greek hospital. *Diagnostic Microbiology and Infectious Disease*. 29(1), 39-41.
- [43] Yang, Q., Zhang, H., Wang, Y., Xu, Z., Zhang, G., Chen, X. (2017) Antimicrobial susceptibilities of aerobic and facultative gram-negative bacilli isolated from Chinese patients with urinary tract infections between 2010 and 2014. *BMC Infectious Diseases*. 17,192-199.
- [44] Zarakolu, P., Hascelik, G., Unal, S. (2006) Antimicrobial susceptibility pattern of nosocomial gram negative pathogens: Results from mystic study in Hacettepe university adult hospital (2000-2004). *Bulletin of Microbiology*. 40, 147-154.
- [45] Bashir, R., Zaib, N., Altaf, I., Saleem, F., Sultana, Q., Naz, S. (2016) Optimization of PCR for rapid detection of CTX-M gene in ESBL producing *Klebsiella pneumoniae* clinical isolates from Punjab, Pakistan. *Malaysian Journal of Microbiology*. 12(5), 365-369.
- [46] Uzun, K., Teke, T., Yavuz, Z.G. (2006) Surveillance of antimicrobial resistance and susceptibility in bacteria isolated from pulmonart critical care. *Tip Araştır Derg*. 4(3), 8-13.
- [47] Kader, A.A., Kumar, A. (2004) Prevalence and antimicrobial susceptibility of extended-spectrum  $\beta$ -lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* in a general hospital in Saudi Arabia. *Saudi Medical Journal*. 25, 570-574.
- [48] Yılmaz, N., Köse, Ş., Ağuş, N., Ece, G., Akkoçlu, G., Kıraklı, C. (2010) Microorganisms isolated from blood cultures of intensive care unit patients, their antimicrobial susceptibility and etiological agents in nosocomial bacteremia. *Ankem Journal*. 24(1), 12-19.
- [49] İnan, N.U., Gurler, N. (2004) Investigation of antibiotic resistance and some virulence factors of *Escherichia coli* strains isolated from children with urinary tract infections. *Ankem Journal*. 18(2), 89-96.
- [50] Barišić, Z., Babić-Erceg, A., Borzić, E., Zoranić, V., Klaiterna, V., Carev, M. (2003) Urinary tract infections in South Croatia: aetiology and antimicrobial resistance. *International Journal of Antimicrobial Agents*. 22, 61-64.
- [51] Sohail, M., Khurshid, M., Saleem, H.G.M., Javed, H., Khan, AA. (2015) Characteristics and antibiotic resistance of urinary tract pathogens isolated from Punjab, Pakistan. *Jundishapur Journal of Microbiology*. 8(7), 1-5.
- [52] Ekim, M., Kuluglu, Z., Aysev, D., Can, S. (1998) Changes in antibiotic sensitivity in urinary tract infection caused by *Escherichia coli*. *Journal of the Turkish Nephrology*. 3 141-144.
- [53] Cetin, E.S., Demirci, M., Kaya, S., Arıdoğan, B.C., Adiloglu, A.K., Goksu, Y., Gonulates, N. (2006) Antibiotic susceptibilities of *Escherichia coli*, *Klebsiella pneumoniae* and Enterobacter strains isolated from blood specimens. *Turkish Microbiology Society Journal*. 36(1), 20-24.
- [54] Kumar, A.R. (2013) Antimicrobial sensitivity pattern of *Klebsiella pneumoniae* isolated from pus from tertiary care hospital and issues related to the rational selection of antimicrobials. *Journal of Chemical Pharmaceutical Research*. 5(11), 326-331.
- [55] Manikandan, C., Amsath, A. (2013) Antibiotic susceptibility of bacterial strains isolated from patients with respiratory tract infections. *International Journal of Pure and Applied Zoology*. 1(1), 61-69.
- [56] Al-Mardeni, R.I., Batarseh, A., Omaish, L., Shraideh, M., Batarseh, B., Unis, N. (2009) Empirical treatment for pediatric urinary tract infection and resistance patterns of uropathogens, in Queen Alias Hospital and Prince A'isha Military Center. *Jordan. Saudi J Kidney Dis Transplant*. 20, 135-9.
- [57] Mathai, E., Chandy, S., Thomas, K., Antoniswamy, B., Joseph, I., Mathai, M., Sorensen, T.L., Holloway, K. (2008) Antimicrobial resistance surveillance among commensal *Escherichia coli* in rural and urban areas in Southern India. *Trop Med Int Health*. 13(1), 415.
- [58] Al-Tawfiq, J.A. (2006) Increasing antibiotic resistance among isolates of *Escherichia coli* recovered from inpatients and outpatients in a Saudi Arabian hospital. *Infectious Control Hospital Epidemiology*. 27, 748-753.
- [59] Paul, S., Bezbaruah, R.L., Roy, M.K., Ghosh, A.C. (1997) Multiple antibiotic resistance (MAR) index and its reversion in *Pseudomonas aeruginosa*. *Letters in Applied Microbiology*. 24, 169-171.



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