



Noninvasive Models to Predict Liver Fibrosis in Patients with Chronic Hepatitis B: A Study from Turkey

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Abstract

Background: Many noninvasive methods, including aspartate amino transaminase (AST)/alanine amino transaminase (ALT) ratio (AAR), AST-to-platelet ratio index (APRI), Bonacini cirrhosis discriminant score (CDS), fibrosis-4 (FIB4) index, and age-platelet index (API), have been described to determine the stage of hepatic fibrosis. However, these methods are developed for patients with chronic hepatitis C (CHC) and produce conflicting results in the prediction of liver fibrosis in patients with chronic hepatitis B (CHB).

Objectives: The aim of this study was to evaluate the relationship between 7 noninvasive models, including AAR, APRI, CDS, API, FIB-4, neutrophil-to-lymphocyte ratio (NLR), and red cell distribution width (RDW)-to-platelet ratio (RPR) in patients with CHB.

Methods: The study population included all patients with CHB, undergoing liver biopsy to determine HBsAg and HBV DNA positivity in more than 6 months.

Results: A total of 2520 treatment-naïve CHB patients from 40 different centers were included in the study. In total, 62.6% of the patients were male, and the mean age was 40.60 ± 12.34 years (minimum, 18 years; maximum, 77 years). The Ishak fibrosis score was ≥ 3 in 29.8% of the patients, indicating significant fibrosis. The mean API, APRI, CDS, NLR, FIB4, and RPR scores in the noninvasive models were significantly different between the groups with significant and low fibrosis ($P < 0.05$). All the noninvasive models (API, APRI, AAR, CDS, NLR, RPR, and FIB4) were found to be significant in the discrimination of cirrhosis ($P < 0.05$). In the multiple logistic regression analysis, CDS, albumin, alkaline phosphatase (ALP), total bilirubin, neutrophil count, NLR, mean platelet volume (MPV), and FIB4 were independent indices for cirrhosis.

Conclusions: In the present study, the role of noninvasive tests in the prediction of liver fibrosis stage and cirrhosis was evaluated in a large cohort of CHB patients. Overall, noninvasive models are gradually becoming more promising. Accordingly, the need for liver biopsy can be reduced with a combination of noninvasive methods in the future.

Keywords: Chronic Hepatitis B, Liver, Fibrosis, Noninvasive Models

1. Background

Approximately one-third of the world's population is infected with hepatitis B virus (HBV), and 350 - 400 million people are chronic hepatitis B surface antigen (HBsAg) car-

riers. The disease has a broad clinical spectrum, ranging from inactive carriers to cirrhosis and hepatocellular carcinoma (HCC). The major aim of treatment is to prevent disease progression to advanced stages, such as cirrhosis and HCC.

Diagnosis of hepatic fibrosis is important in the selection of treatment (1). Liver biopsy is the gold standard for the evaluation of hepatic fibrosis stage (2). Although percutaneous liver biopsy is generally considered a safe procedure, it is costly and may increase the risk of complications (3). Biopsy can only represent 1:50.000 of the liver, and its accuracy may be influenced by the techniques and methodological interventions. For instance, fibrosis stage cannot be classified in 20% of the patients (4, 5).

Low compliance of patients due to the invasive nature of biopsy, contraindication of liver biopsy in hemostasis disorders, and limited use of liver biopsy during follow-up are the limitations of the procedure (3, 6, 7). The other limitations include the need for static information about the time of liver biopsy for fibrosis, variability of histological findings from the right and left hepatic lobes (considering the biopsy performance), frequent fragmentation of biopsy specimens during biopsy of cirrhotic patients, and finally a fibrosis level lower than expected (8-10).

Many noninvasive methods, including aspartate amino transaminase (AST)/alanine amino transaminase (ALT) ratio (AAR), AST-to-platelet ratio index (APRI), Bonacini cirrhosis discriminant score (CDS), fibrosis-4 (FIB4) index, and age-platelet index (API), have been described to evaluate the degree of hepatic fibrosis (11-15). However, these methods are developed for patients with chronic hepatitis C (CHC) and produce conflicting results in the prediction of liver fibrosis in patients with chronic hepatitis B (CHB) (16). CHC and CHB are different in terms of virological characteristics, histological changes in the liver, and triggering mechanisms for fibrosis (17). Therefore, it is necessary to evaluate the use of these parameters in patients with CHB.

2. Objectives

The aim of this study was to evaluate the relationship between 7 noninvasive models, including AAR, APRI, CDS, API, FIB-4, neutrophil-to-lymphocyte ratio (NLR), and red cell distribution width (RDW)-to-platelet ratio (RPR) in patients with CHB.

3. Methods

3.1. Study Population

The present research was designed as a retrospective multicenter study, performed at infectious diseases and clinical microbiology and gastroenterology clinics. All patients with CHB undergoing liver biopsy for HBsAg and HBV DNA analysis in more than 6 months, between January 2011 and December 2015, were included in the study. On the

other hand, the exclusion criteria were as follows: 1) regular use of alcohol (> 20 g in a week); 2) use of anti-inflammatory drugs; 3) decompensated liver disease, renal failure, diabetes mellitus, chronic inflammatory diseases (eg, ulcerative colitis, Crohn's disease, and rheumatoid arthritis), coinfection (eg, HIV, HCV, and HDV), and HCC; 4) other malignancies metabolic and autoimmune liver diseases; and 5) history of liver surgery or transplantation. The medical records of patients undergoing liver biopsy for CHB were evaluated retrospectively. The study was approved by Dumlupinar University clinical research ethics committee.

3.2. Laboratory Assessment

The patients' leukocyte count, neutrophil count, lymphocyte, platelet count, hemoglobin, RDW, mean corpuscular volume (MCV), and mean platelet volume (MPV) were recorded, based on the complete blood cell count before biopsy. AST, ALT, albumin, total bilirubin, gamma-glutamyl transferase (GGT), and alkaline phosphatase (ALP) were recorded among biochemical parameters, and the international normalized ratio (INR) was reported among coagulation parameters. HBsAg, anti-HB antibody, HBeAg, anti-HBe antibody, HBV DNA, delta-antigen, anti-delta, anti-HIV, and anti-HCV antibodies were also evaluated in viral serological tests.

3.3. Histological Assessment

Liver biopsy specimens of the patients were assessed by pathologists in the follow-up location. Liver biopsy specimens, which were assessed according to the histological activity index (HAI) and Ishak fibrosis index, were included in the study (7). The specimens with fibrosis scores of 0 - 2 were classified as low fibrosis, scores ≥ 3 were classified as significant fibrosis, and scores of 5 - 6 were defined as cirrhosis. Noninvasive models were compared between patients with low and significant fibrosis, as well as cirrhotic and noncirrhotic patients.

3.4. Noninvasive Models

Among the noninvasive models, AAR, API, CDS, FIB4, and APRI were measured for each patient, using different formulas: AAR (De Ritis ratio): AST/ALT; API: age + platelet count (age: < 30 years, 0; 30 - 39 years, 1; 40 - 49 years, 2; 50 - 59 years, 3; 60 - 69 years, 4; ≥ 70 years, 5); platelet count $\times 10^9/L$ (≥ 225 , 0; 200 - 224, 1; 175 - 199, 2; 150 - 174, 3; 125 - 149, 4; < 125, 5); CDS: platelet count + ALT/AST ratio + INR (platelet count $\times 10^9/L$ > 340, 0; 280 - 339, 1; 220 - 279, 2; 160 - 219, 3; 100 - 159, 4; 40 - 99, 5; < 40, 6) (ALT/AST ratio > 1.7, 0; 1.2 - 1.7, 1; 0.6 - 1.19, 2; < 0.6, 3) (INR < 1.1, 0; 1.1 - 1.4, 1; > 1.4, 2); APRI: $([AST/ULN]/platelet\ count\ [\times 10^9/L]) \times 100$; and FIB4: $[age \times$

AST (U/L)/platelet count ($10^9/L$) \times ALT (U/L)/2] (11-15). Moreover, NLR was calculated by dividing the neutrophil count to lymphocyte count in the complete blood cell count, and RPR was calculated by dividing RDW to platelet count (17, 18).

3.5. Statistical Analysis

The number cruncher statistical system (NCSS 2007; Kaysville, Utah, USA) was used for the statistical analysis. For the comparison of descriptive results, mean, standard deviation, median, frequency, ratio, minimum, and maximum values were calculated. In addition, regarding quantitative data, student t test was used for intergroup comparisons of parameters with normal distribution, while Mann-Whitney U test was applied for the intergroup comparisons of parameters without a normal distribution. Diagnosis screening, including sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), and receiver operating curve (ROC) analyses, were used to determine the cut-off values for the parameters. Multivariate logistic regression analysis was used to determine the risk factors for fibrosis and cirrhosis. The significance level was set at $P < 0.05$.

4. Results

A total of 2520 treatment-naive CHB patients from 40 different centers were included in the study. Overall, 62.6% of the patients were male, and the mean age was 40.60 ± 12.34 years (minimum, 18 years; maximum, 77 years). The Ishak fibrosis score was ≥ 3 in 29.8% of the patients, indicating significant fibrosis (Table 1). According to the comparisons, while no significant difference was found regarding hemoglobin, lymphocyte, RDW, leukocyte, or MPV ($P > 0.05$), a significant difference was found in the platelet count, albumin, neutrophil count, AST, ALT, INR, GGT, ALP, total bilirubin, and MCV between the groups with low and significant fibrosis ($P < 0.05$). The prevalence of significant fibrosis was significantly higher in male patients and those with advanced age ($P < 0.05$).

While albumin level, platelet count, and neutrophil count were significantly lower in the group with significant fibrosis, age, AST, ALT, INR, GGT, ALP, total bilirubin, and MCV were significantly lower in the group with significant fibrosis. API, AST, ALT, INR, albumin, GGT, ALP, and MCV were independent variables for the diagnosis of significant fibrosis in the multiple logistic regression analysis. Two most important independent variables associated with significant fibrosis were INR (odds ratio, 5.234; 95% CI, 2.73 - 10.04) and API (odds ratio, 1.668; 95% CI, 1.28 - 2.16). However, no significant difference was found regarding the mean AAR between the groups ($P = 0.524$; Table 2).

Table 1. The Demographic Characteristics of the Patients

	Min-Max (Median)
Age	18 - 77 (40)
Female	942 (27.4)
Male	1578 (62.6)
Total bilirubin, mg/dL	0.1 - 3.5 (0.7)
AST, U/L	5 - 599 (35)
ALT, U/L	6 - 1053 (52)
Albumin, g/dL	2-5.6 (4.3)
GGT, U/L	1 - 670 (26)
ALP, U/L	10 - 511 (78)
Neutrophil, $10^3/\mu L$	0.8 - 11.6 (3.7)
Lymphocyte, $10^3/\mu L$	0.2 - 7 (2.1)
Hb, g/dL	7.5 - 18.8 (14.6)
RDW	6.3 - 47.7 (13.5)
MCV	35.8 - 120.7 (87.8)
MPV	5.5 - 22.8 (9.2)
Platelet count, $10^9/L$	56 - 640 (215)
INR	0.4 - 3.1 (1)
HBeAg negative	1966 (78)
HBeAg positive	554 (22)
HBV DNA $\times 10^4$, IU/mL	0.2 - 627924 (24.9)
HAI	0 - 18 (6)
F0	345 (13.7)
F1	612 (24.3)
F2	810 (32.1)
F3	452 (17.9)
F4	187 (7.4)
F5	96 (3.8)
F6	18 (0.7)

Among the noninvasive models, the mean API, APRI, CDS, NLR, FIB4, and RPR were significantly different between the groups with significant and low fibrosis. The ROC curves of these models are presented in Figure 1.

The area under ROC (AUROC) for the noninvasive models of API, APRI, CDS, NLR, FIB4, and RPR were 0.629, 0.688, 0.609, 0.470, 0.681, and 0.596, respectively among CHB patients. The cut-off point for NLR in discrimination of low and significant fibrosis was ≤ 1.63 . Moreover, the cut-off points for API, CDS, APRI, FIB4, and RPR, according to the fibrosis scores, were $> 3, 5, 0.54, 1.19,$ and 0.07 , respectively. The optimal cut-off point, sensitivity, specificity, PPV, and NPV of 6 noninvasive models, significant in the discrimi-

Table 2. Group Comparisons and Logistic Regression Analysis of Patients with CHB According to Fibrosis^a

	Group Comparisons			Logistic Regression Analysis	
	Low Fibrosis (n, 1767)	Significant Fibrosis (n, 753)	P Value	Univariate OR (95% CI)	Multivariate OR (95% CI)
Age	39.29 ± 11.92	43.67 ± 12.76	0.001 ^b	1.029 (1.022 - 1.037)	
Gender (male)	1072 (60.7)	506 (67.2)	0.002 ^b	1.328 (1.100 - 1.589)	
Platelet count	228.03 ± 62.76	206.98 ± 68.46	0.001 ^b	0.995 (0.993 - 0.996)	
AST	31 (23 - 48)	48 (31 - 89)	0.001 ^b	1.011 (1.009 - 1.013)	1.005 (1.001 - 1.010)
ALT	45.5 (24 - 80)	67 (40 - 128)	0.001 ^b	1.004 (1.003 - 1.005)	0.998 (0.995 - 1.000)
INR	1 (1 - 1.1)	1.1 (1 - 1.2)	0.001 ^b	13.825 (8.068 - 23.691)	5.234 (2.729 - 10.040)
Albumin	4.29 ± 0.43	4.16 ± 0.47	0.001 ^b	0.534 (0.439 - 0.649)	0.719 (0.557 - 0.927)
GGT	24 (16 - 36)	33 (21 - 54)	0.001 ^b	1.013 (1.009 - 1.016)	1.003 (1.001 - 1.006)
ALP	75 (60 - 91)	85 (67 - 109)	0.001 ^b	1.009 (1.007 - 1.012)	1.006 (1.003 - 1.009)
Total bilirubin	0.6 (0.4 - 0.9)	0.7 (0.5 - 1)	0.001 ^b	1.782 (1.446 - 2.197)	
HBeAg positive	375 (21.2)	181 (24.1)	0.112	1.178 (0.962 - 1.443)	
HBV DNA × 10 ⁴	10.2 (1.1 - 344.3)	174.4 (8.3 - 1685)	0.001 ^b	1.000 (1.000 - 1.000)	
Hb	14.42 ± 1.72	14.41 ± 1.66	0.855	0.995 (0.947 - 1.047)	
Leukocyte	6.8 (5.6 - 8)	6.6 (5.5 - 7.8)	0.051	0.964 (0.919 - 1.010)	
Neutrophil	3.8 (3 - 4.8)	3.5 (2.8 - 4.5)	0.001 ^b	0.892 (0.837 - 0.950)	
Lymphocyte	2.1 (1.7 - 2.6)	2.1 (1.7 - 2.6)	0.452	0.957 (0.845 - 1.085)	
RDW	13.4 (12.8 - 14.4)	13.7 (12.8 - 14.7)	0.061	0.989 (0.955 - 1.025)	
MPV	9.31 ± 1.74	9.42 ± 1.83	0.164	1.035 (0.986 - 1.085)	
MCV	87.3 (84.1 - 90.4)	88.8 (85.3 - 91.9)	0.001 ^b	1.051 (1.035 - 1.067)	1.017 (1.00 - 1.036)
API	2 (1 - 4)	3 (2 - 5)	0.001 ^b	1.272 (1.220 - 1.327)	1.668 (1.286 - 2.164)
APRI	0.4 (0.2 - 0.6)	0.6 (0.4 - 1.1)	0.001 ^b	2.403 (2.063 - 2.799)	
NLR	1.8 (1.4 - 2.3)	1.7 (1.3 - 2.3)	0.021 ^c	0.914 (0.834 - 1.002)	
RPR	0.06 (0.05 - 0.07)	0.07 (0.06 - 0.09)	0.001 ^b	2.452 (1.985 - 3.027)	
FIB4	0.8 (0.6 - 1.2)	1.3 (0.8 - 2.1)	0.001 ^b	2.040 (1.827 - 2.276)	
CDS	4.85 ± 1.42	5.41 ± 1.46	0.001 ^b	1.319 (1.240 - 1.404)	
AAR	0.7 (0.6 - 1)	0.7 (0.6 - 0.9)	0.524	1.050 (0.857 - 1.287)	

^aValues are presented as mean ± SD, No. (%) or median (25 - 75 quartiles).

^bP < 0.01.

^cP < 0.05.

nation of low and significant fibrosis (API, APRI, CDS, NLR, FIB4, and RPR), are presented in [Table 3](#).

The comparison of groups with and without cirrhosis indicated that hemoglobin, leukocyte, and lymphocyte levels were not significantly different between the groups ($P > 0.05$). However, there was a significant difference regarding the platelet count, albumin level, neutrophil count, AST, ALT, INR, GGT, ALP, total bilirubin, HAI, MCV, and RDW between the groups ($P < 0.05$). All the noninvasive models (API, APRI, AAR, CDS, NLR, RPR, and FIB4) were significant in the discrimination of cirrhosis ($P < 0.05$). The ROC curves of noninvasive models for the prediction of cir-

rhosis are shown in [Figure 2](#). The prevalence of cirrhosis was significantly higher in male patients and those with advanced age ($P < 0.05$; [Table 4](#)).

While albumin, platelet count, and neutrophil count were found to be lower in the cirrhotic group, compared to the noncirrhotic group, age, AST, ALT, INR, GGT, ALP, total bilirubin, and MCV were significantly higher in the cirrhotic group ($P \leq 0.05$). In the multiple logistic regression analysis, CDS, albumin, ALP, total bilirubin, neutrophil count, NLR, MPV, and FIB4 were independent risk factors for cirrhosis. The most important independent factor for cirrhosis was FIB4 (odds ratio, 2.349; 95% CI, 1.31 - 4.199), fol-

Table 3. The Sensitivity, Specificity, Predictive Values, and AUROC of Noninvasive Models (API, CDS, APRI, NLR, FIB4, and RPR) for the Prediction of Significant Fibrosis

	Diagnostic Test					ROC Curve			P Value
	Cut-Off	Sensitivity	Specificity	PPV	NPV	Area	SE	95% CI	
API	> 3	48.87	70.68	41.49	76.47	0.629	0.013	0.604 - 0.653	0.001 ^a
CDS	> 5	48.47	67.80	39.04	75.57	0.609	0.013	0.584 - 0.633	0.001 ^a
APRI	> 0.54	56.57	71.41	45.71	79.45	0.688	0.012	0.667 - 0.713	0.001 ^a
NLR	≤ 1.63	46.75	59.72	33.05	72.50	0.470	0.013	0.445 - 0.496	0.022 ^b
FIB4	> 1.19	55.25	74.46	47.93	79.64	0.681	0.012	0.658 - 0.706	0.001 ^a
RPR	> 0.07	40.33	75.47	40.78	75.13	0.596	0.013	0.571 - 0.622	0.001 ^a

^aP < 0.01.^bP < 0.05.**Table 4.** Group Comparisons and Logistic Regression Analysis of Patients with CHB for the Prediction of Cirrhosis^a

	Group Comparisons			Logistic Regression Analysis	
	Noncirrhotic (n, 2406)	Cirrhotic (n, 114)	P Value	Univariate OR (95% CI)	Multivariate OR (95% CI)
Age	40.20 ± 12.16	48.96 ± 13.17	0.001 ^b	1.057 (1.041 - 1.074)	
Gender (male)	1496 (62.2)	82 (71.9)	0.035 ^c	1.559 (1.027 - 2.365)	
Platelet count	223.95 ± 64.37	175.25 ± 65.91	0.001 ^b	0.985 (0.981 - 0.989)	
AST	34 (24 - 57)	59.5 (37 - 101)	0.001 ^b	1.007 (1.004 - 1.009)	
ALT	51 (27 - 93)	68 (41.8 - 133)	0.001 ^b	1.002 (1.001 - 1.004)	
INR	1 (1 - 1.1)	1.1 (1 - 1.2)	0.001 ^b	2.020 (1.094 - 3.731)	
Albumin	4.26 ± 0.43	3.92 ± 0.53	0.001 ^b	0.248 (0.173 - 0.354)	0.411 (0.243 - 0.693)
GGT	25 (17 - 41)	41 (26 - 70)	0.001 ^b	1.006 (1.003 - 1.009)	
ALP	77.5 (62 - 95)	84 (67 - 115.5)	0.007 ^b	1.008 (1.004 - 1.011)	1.005 (1.000 - 1.010)
Total bilirubin	0.6 (0.5 - 0.9)	0.9 (0.7 - 1.2)	0.001 ^b	3.019 (2.156 - 4.227)	1.612 (0.934 - 2.782)
HBeAg positive	525 (21.8)	31 (27.2)	0.176	1.339 (0.876 - 2.045)	
HBV DNA × 10 ⁴	21.2 (1.6 - 558.6)	221.7 (20 - 2492.5)	0.001 ^b	1.000 (1.000 - 1.000)	
Hb	14.42 ± 1.70	14.30 ± 1.82	0.441	0.958 (0.859 - 1.068)	
Leukocyte	6.7 (5.6 - 8)	6.6 (5.5 - 7.6)	0.347	0.946 (0.851 - 1.052)	
Neutrophil	3.7 (2.9 - 4.7)	3.4 (2.7 - 4.5)	0.029 ^c	0.879 (0.761 - 1.016)	1.252 (1.051 - 1.491)
Lymphocyte	2.1 (1.7 - 2.6)	2.1 (1.7 - 2.7)	0.564	1.101 (0.843 - 1.438)	
RDW	13.5 (12.8 - 14.5)	13.9 (13.1 - 14.9)	0.009 ^b	1.027 (0.966 - 1.091)	
MCV	87.7 (84.3 - 90.9)	89 (85.8 - 92.4)	0.007 ^b	1.039 (1.006 - 1.074)	
MPV	9.32 ± 1.76	9.77 ± 1.89	0.009 ^b	1.129 (1.031 - 1.238)	1.220 (1.055 - 1.412)
FIB4	0.9 (0.6 - 1.4)	1.9 (1.3 - 3.1)	0.001 ^b	1.713 (1.538 - 1.908)	2.349 (1.314 - 4.199)
API	3 (1 - 4)	5 (3 - 7)	0.001 ^b	1.579 (1.452 - 1.718)	
APRI	0.4 (0.3 - 0.7)	0.9 (0.5 - 1.6)	0.001 ^b	1.692 (1.467 - 1.952)	
AAR	0.7 (0.6 - 1)	0.8 (0.6 - 1)	0.020 ^c	1.475 (1.085 - 2.005)	
CDS	4.97 ± 1.45	5.86 ± 1.39	0.001 ^b	1.564 (1.361 - 1.797)	1.259 (1.056 - 1.501)
NLR	1.8 (1.4 - 2.3)	1.6 (1.3 - 2.2)	0.046 ^c	0.895 (0.719 - 1.116)	2.018 (1.162 - 3.503)
RPR	0.06 (0.05 - 0.08)	0.08 (0.07 - 0.11)	0.001 ^b	5.031 (3.420 - 7.400)	

^aValues are presented as mean ± SD, No. (%), or median (25 - 75 quartiles).^bP < 0.01.^cP < 0.05.

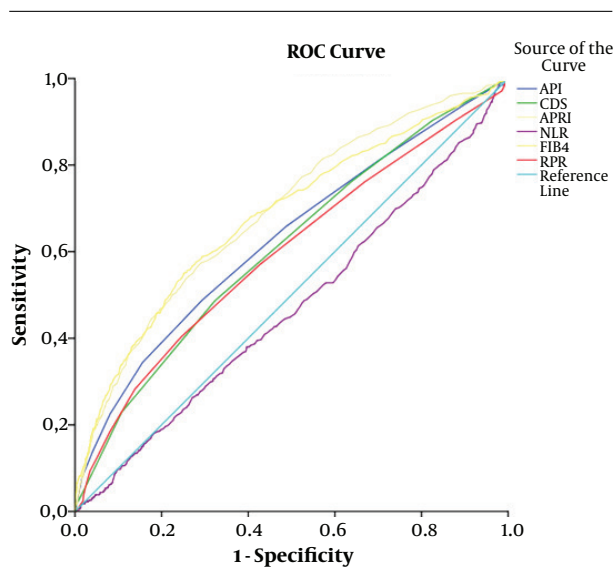


Figure 1. The ROC of Noninvasive Models (API, CDS, APRI, NLR, FIB4, and RPR) in CHB Patients

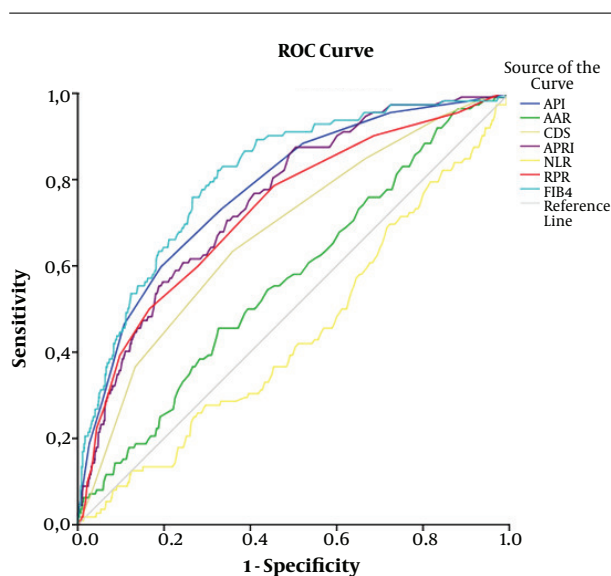


Figure 2. The ROC of Noninvasive Models (API, AAR, CDS, APRI, NLR, FIB4, and RPR) in Cirrhotic Patients

lowed by NLR (odds ratio, 2.108; 95% CI, 1.162 - 3.503; Table 4). The cut-off points for CDS, API, AAR, APRI, FIB4, and RPR in cirrhotic patients were 5, 4, 0.86, 0.4, 1.17, and > 0.08 , respectively. Moreover, the cut-off point for NLR in cirrhotic patients was ≤ 1.62 . The optimum cut-off point, sensitivity, specificity, PPV, and NPV of 7 noninvasive models (API, APRI, CDS, NLR, FIB4, and RPR) in the prediction of cirrhosis are shown in Table 5.

5. Discussion

Based on the findings, 6 out of 7 noninvasive models could successfully identify fibrosis in CHB patients without biopsy. The APRI model, which was first investigated by Wai et al. in the staging of fibrosis among CHB patients, showed the most accurate diagnostic value and most powerful predictive value for fibrosis among the models in this study (19). The AUROC of APRI in the prediction of significant fibrosis was 0.688; this finding seems to be consistent with the data reported in the literature (20-24). The NPV of APRI in the prediction of fibrosis and cirrhosis was $> 90\%$ in a study by Guzelbulut et al. and 92% in a study by Celikbilek et al. (20, 22). In the present study, the NPVs of APRI for fibrosis and cirrhosis were 79.45% and 98.84%, respectively. These findings show that APRI is an important indicator, which can exclude fibrosis and cirrhosis and can be used accordingly in CHB patients.

The cut-off AUROC of FIB4, which is another index used to evaluate liver fibrosis, was 1.19; the sensitivity and specificity of FIB4 for significant fibrosis were 55.25% and 74.46%, respectively. This cut-off value seems to be consistent with the values reported in the literature (21, 23). In a meta-analysis by Xiao et al. the sensitivity and specificity of FIB4 in the prediction of cirrhosis were 87.4% and 64.7%, respectively (25). In the present study, the sensitivity and specificity of FIB4 in the prediction of cirrhosis were 81.5% and 67.3%, respectively, based on the AUROC. These findings are consistent with the results obtained by Xiao and colleagues. Moreover, in the present study, the NPVs of FIB4 to exclude significant fibrosis and cirrhosis were 79.64% and 98.7%, respectively. Consequently, FIB4 seems to be an important noninvasive assessment tool, which can be used to exclude significant fibrosis, especially cirrhosis.

Lymphocytes play a regulatory role in immune responses and have been shown to influence liver fibrosis in CHB patients (22). NLR is an inexpensive noninvasive model, showing the host's immune responses. Simply, it can be calculated via complete blood cell count parameters. NLR was found to be useful in the evaluation of HCC prognosis and discrimination of nonalcoholic steatohepatitis and significant fibrosis (26, 27). In various studies, NLR has been shown to efficiently identify the stage of liver

Table 5. The Sensitivity, Specificity, PPV, NPV, and AUROC of Noninvasive Models (API, AAR, CDS, APRI, NLR, FIB4, and RPR) for the Prediction of Cirrhosis

	Diagnostic Test					ROC Curve			P Value
	Cut-Off	Sensitivity	Specificity	PPV	NPV	Area	SE	95% CI	
API	> 4	59.65	80.61	12.71	97.69	0.775	0.022	0.758 - 0.791	0.001 ^a
AAR	> 0.86	44.74	67.12	6.05	96.25	0.564	0.027	0.545 - 0.584	0.017 ^b
CDS	> 5	64.04	64.22	7.81	97.42	0.679	0.025	0.661 - 0.698	0.001 ^a
APRI	> 0.4	87.72	49.56	7.60	98.84	0.752	0.022	0.735 - 0.769	0.001 ^a
NLR	≤ 1.62	54.39	59.07	5.92	96.47	0.555	0.028	0.536 - 0.575	0.047 ^b
FIB4	> 1.17	81.58	67.37	10.58	98.72	0.804	0.021	0.788 - 0.819	0.001 ^a
RPR	> 0.08	50.00	83.42	12.53	97.23	0.728	0.025	0.710 - 0.745	0.001 ^a

^a P < 0.01.^b P < 0.05.

fibrosis among CHB patients and can be used to demonstrate liver fibrosis (18, 28). On the contrary, some studies have indicated no relationship between NLR and cirrhosis and liver fibrosis (22). In the present study, NLR could be used in the discrimination of both significant fibrosis and cirrhosis. Analysis of the relationships, especially between NLR and cirrhosis, was performed using multiple regression analysis, and NLR was determined to be an independent factor for cirrhosis. NLR also showed the greatest value effect, following FIB4 among multiple factors for cirrhosis. However, the efficiency of NLR in the identification of liver fibrosis should be supported in future studies.

RDW is an indicator of variability in erythrocyte volume in circulation. In addition, it is used for the differential diagnosis of anemia (16). The increase in RDW has been shown to be proportional to the severity of liver damage. Thrombocytopenia is one of the well-known markers of severe liver injury. RPR is the ratio of RDW to platelet count and can be calculated according to the complete blood cell count parameters, such as NLR. In the literature, some studies have used RPR in staging of liver fibrosis (16, 29, 30). Chen et al. reported RPR as a cost-effective and easily measurable method and found it to be superior to AAR, FIB4, and APRI in the prediction of cirrhosis (16). In the present study, AUROC of RPR for significant fibrosis and cirrhosis was 0.596 and 0.728, respectively. According to the present findings, RPR was a more valuable parameter in the diagnosis of significant fibrosis in comparison with cirrhosis among CHB patients.

CDS is another model in which liver fibrosis is evaluated noninvasively. For the first time, in a study by Bonacini et al. a positive correlation was found between fibrosis score and CDS in CHC patients. In their study, the cut-off value, sensitivity, and specificity of CDS for advanced fibrosis or cirrhosis were > 8, 46%, and 98%, respectively (11). Overall, the number of studies indicating a relationship between CDS and fibrosis staging is limited in CHB patients. While CDS was insufficient in the diagnosis of liver fibro-

sis in some studies, it could be used to diagnose fibrosis in several other studies (21, 24, 30). In the present study, the cut-off value and NPV of CDS for low and significant fibrosis in cirrhotic and noncirrhotic patients were > 5, 75.57%, and 97.42%, respectively.

According to the present results, CDS is efficient in the discrimination of low fibrosis, significant fibrosis, and cirrhosis. However, since a higher NPV was obtained in the discrimination of cirrhosis, it is more successful in the evaluation of cirrhosis. Since no previous studies in the literature have evaluated the use of CDS in the discrimination of cirrhosis among CHB patients, the present findings can shed light on future studies.

For the first time, Poynard et al. evaluated API as a model for fibrosis in CHC patients (12). This model has been shown to be a good predictive index for liver fibrosis stage in many studies (24, 30). In the present study, AUROC for API was 0.629, which is consistent with the literature. On the contrary, Erdogan et al. reported that API is ineffective in the discrimination of significant fibrosis (21). Moreover, in an extensive meta-analysis, API was a successful noninvasive model for cirrhosis and NPV was 99.1% (25). Moreover, in the present study, the NPV of API was 97.69% for cirrhosis and API was an effective model.

In addition, AAR is a common index for identifying the underlying liver disease. AAR is generally > 2 in alcoholic liver disease and < 1 in chronic hepatitis and chronic cholestatic syndrome (24). In the present study, AAR was ineffective in differentiation of low fibrosis from significant fibrosis. There are studies supporting our results in the literature (21, 24, 31). In some studies, AAR is a parameter, used to identify cirrhosis rather than fibrosis (23). Similarly, we showed that AAR is an index for cirrhosis (Table 4).

INR is the most important indirect and noninvasive index for the prediction of hepatic cirrhosis. ALT, AST, and platelet count are correlated with liver cell death. In liver damage, an increase in PT/INR ratio and a decrease in albumin level have been observed (31). In a study on the

role of laboratory tests in the identification of fibrosis in CHB patients, platelet count, AST, ALT, GGT, ALP, total bilirubin, albumin, and INR were compared between patients with liver fibrosis stages 0 - 1 and 2 - 4; these parameters were found to be significantly different between the groups with respect to fibrosis stage (32). In another study, age, INR, AST, and GGT were associated with liver fibrosis stage (21). In addition, in our study, the association between fibrosis and parameters such as age, albumin level, platelet count, HBV DNA, neutrophil count, AST, ALT, INR, GGT, ALP, total bilirubin, and MCV was investigated. AST, ALT, INR, albumin, GGT, ALP, and MCV were independent predictors of fibrosis in the regression analysis.

In patients with significant liver fibrosis, platelet count decreases due to reduced thrombopoietin production and increased platelet destruction (14). In this regard, conflicting results have been obtained in studies investigating the association between platelet count and fibrosis (2, 8, 14, 15, 21, 22). In addition, in our study, while platelet count was negatively correlated with fibrosis stage in the univariate analysis, it was not an independent parameter in the regression analysis. Similarly, the association between platelet count and cirrhosis was investigated. The platelet count was valuable in identifying cirrhosis in the univariate analysis, whereas it was not significant in the regression analysis. Based on the analyses, further clinical studies are required to clarify the efficiency of platelet count in the identification of fibrosis or cirrhosis alone.

The present study included the largest cohort of patients and investigated all noninvasive models in order to examine liver fibrosis stage and cirrhosis in the literature. The most important limitation of this study is the retrospective design; therefore, assessment by different pathologists was required for fibrosis staging. On the other hand, use of a similar method by all pathologists and high-quality standards of Ishak system are the strengths of this study, despite its retrospective design. Additionally, inclusion of patients from different centers is important in representing the general population. Sharing data of one center in relevant studies is reported as limitation of the study and it is stated that data obtained should also be validated by multicenter studies (19, 31). The present study is also important in substantiating this validation.

In conclusion, in the present study, the role of noninvasive tests in the prediction of liver fibrosis stage and cirrhosis was evaluated in a large cohort of CHB patients. Six out of seven noninvasive models for fibrosis stage were efficient in the prediction of significant liver fibrosis. AAR was an insufficient model in staging fibrosis. All 7 noninvasive models were found to be predictive of cirrhosis. FIB4 and APRI were the noninvasive models with the highest diagnostic accuracies for fibrosis stage, while FIB4 and API

were the noninvasive models with the highest diagnostic accuracy for cirrhosis. Liver biopsy is currently the gold standard for staging hepatic fibrosis; nevertheless, its status can change in the future. Noninvasive models are gradually becoming more promising, and the need for liver biopsy can be reduced with combinations of noninvasive methods in the future.

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Footnotes

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