

Minireview

Molecular mechanisms of action of hesperidin in cancer: Recent trends and advancements

Vaishali Aggarwal¹, Hardeep S Tuli², Falak Thakral², Paavan Singhal², Diwakar Aggarwal², Saumya Srivastava³, Anjana Pandey³, Katrin Sak⁴, Mehmet Varol⁵, Md. Asaduzzaman Khan⁶ and Gautam Sethi⁷ 

¹Department of Histopathology, Post Graduate Institute of Medical Education and Research (PGIMER), Chandigarh 160012, India;

²Department of Biotechnology, Maharishi Markandeshwar (Deemed to be University), Ambala 133207, India; ³Department of Biotechnology, MNNIT Allahabad, Prayagraj 211004, India; ⁴Praeventio, NGO, Tartu 50407, Estonia; ⁵Department of Molecular Biology and Genetics, Faculty of Science, Mugla Sitki Kocman University, Mugla TR48000, Turkey; ⁶The Research Center for Preclinical Medicine, Southwest Medical University, Luzhou 646000, China; ⁷Department of Pharmacology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore 117600, Singapore

Corresponding authors: Gautam Sethi. Email: phcgs@nus.edu.sg;

Hardeep S Tuli. Email: hardeep.biotech@gmail.com

Impact statement

Experimental findings from numerous studies have demonstrated the anticancer effects of hesperidin (Hesp) to be associated with anti-oxidant and anti-inflammatory activities along with its potential role in inhibiting the tumor cell metastasis and angiogenesis. Additionally, Hesp can also reverse drug resistance of cancer cells, which make it a promising candidate to be used in combination with existing anti-cancer drugs. This review will be helpful for upcoming researchers and scientific community to find out complete capsular package about cancer drug targets of Hesp and its role in modulating various important hallmarks of cancer.

Abstract

Hesperidin belongs to flavanones class of flavonoids and is known to possess broad-spectrum applicability to prevent dreadful diseases such as cardiovascular disease, neurodegeneration, and cancer. The reported anticancer effects of hesperidin have been found to be associated with its anti-oxidant and anti-inflammatory activities. Hesperidin interacts with numerous recognized cellular targets and inhibits cancer cell proliferation by inducing apoptosis and cell cycle arrest. In addition, evidence has suggested its promising role in inhibiting tumor cell metastasis, angiogenesis, and chemoresistance. The present mini-review highlights the ongoing development to identify hesperidin targets in cancer. Furthermore, the potential of nano technology-based hesperidin combinations and delivery systems will also be discussed. Overall, this review highlights all the possible molecular targets affected by hesperidin in tumor cells on a single platform.

Keywords: Hesperidin, anticancer, synergistic effect, nanotechnology

Experimental Biology and Medicine 2020; 0: 1–12. DOI: 10.1177/1535370220903671

Introduction

Citrus fruits, such as oranges, tangerines, lemons, and limes, are well known for their health-promoting and chemopreventive properties.¹ One of the most important bioactive components which elicits promising anti-cancer potential is a flavonoid hesperidin (Hesp), also known as hesperetin 7-rutinoside.² Hesp was first isolated from orange peels in 1828 and was one of the two compounds formerly erroneously named as “vitamin P.”³ This polyphenolic glycoside can be abundantly found in various citrus fruits.

For instance, sweet oranges contain about 200–600 mg/l of Hesp, whereas the total amount of this molecule varies within 50–850 mg/l in clementines, 8.1–460 mg/l in mandarins, 38–410 mg/l in lime and lemon juice, and 20–170 mg/l in grapefruit juice.¹ Although the bioavailability of dietary flavonoids is known to be limited, the low micromolar levels of Hesp (around 1 μ M range) have been detected in the blood serum for 5–7 h after the intake of citrus juice,^{3,4} probably high enough to exert its health-promoting activities in the human body.

Hesp belongs to the flavanone family of bioflavonoids. Differently from some other common groups of flavonoids, such as flavones, flavonols or isoflavones, flavanones lack the double bond between the positions 2 and 3 in the C ring of general flavonoid structure.¹ Hesp has been shown to exert a broad range of pharmacological activities, being a potent anti-oxidant, anti-inflammatory, anti-atherosclerotic, cardioprotective, neuroprotective, anti-allergic, anti-viral, anti-microbial, and anti-cancer compound.^{1,2,4,5} Its role in protection against malignant transformation and progression has been described in multiple preclinical studies, acting through diverse cellular signalling pathways.^{5,6} Indeed, Hesp can affect diverse molecular targets involved in survival, division, and death mechanisms of tumor cells.^{5,6}

Cancer is the second leading cause of death worldwide, just behind cardiovascular diseases.⁷ In fact, there was an estimated 18.1 million new cancer cases and 9.6 million cancer deaths in 2018 around the world.⁸ Due to the continuously increasing global prevalence of malignancies, novel efficient therapeutics and treatment strategies are highly needed. Application of safe natural compounds with strong anticancer properties, such as Hesp, may open new avenues in cancer treatment.⁹⁻¹¹ Therefore, different anti-tumor mechanisms of this attractive dietary bioflavonoid, i.e. antioxidant, anti-inflammatory, anti-proliferative, anti-angiogenic, anti-invasive, anti-metastatic and, pro-apoptotic properties, are summarized in this review. The dataset presented in this article may be valuable for initiating clinical trials in patients suffering from different cancers, either alone or in combination with traditional therapies.

Chemistry and synthetic preview of hesperidin

On the chemical perspective, hesperidin structure harbors an aglycone known as methyl eriodictyol (hesperetin) bonded to rutinose.¹² Hesp contains a glycoside moiety which is a disaccharide (glucose and rhamnose) which is present in two isomeric forms, i.e. neohesperidose and rutinose.¹² Neohesperidose is chemically known as 2-O- α -L-Rhamnopyranosyl-D-glucopyranose. It is reported to be present in citrus fruits in the form of hesperetin 7-O-neohesperidoside.¹³ Rutinose on the other hand, a disaccharide, usually obtained from herbal sources is chemically 6-O-(α -L-Rhamnosyl)-D-glucose or 6-O-(α -L-Rhamnopyranosyl)-D-glucopyranose.^{12,14} These disaccharide moieties are responsible for the bitterness of citrus bioflavonoids. Of the disaccharides, the taste of citrus fruits is tasteless because of the presence of rutosides moiety, while the neohesperidosides moiety is responsible for the bitter taste. Hesp is commonly found in neohesperidosides in the form of grapefruit (bitter) and in rutoside in the form of orange (non-bitter).^{12,14,15} In the chemical Hesp skeleton, hesperetin (aglycone structure) is bonded to glucose and rhamnose is further bonded through glucose moiety to this structure.¹²

In hesperetin, a bioflavonoid, aglycone structure is (S)-2,3-dihydro-5,7-dihydroxy-2-(3-hydroxy-4-methoxyphenyl)-4H-1-benzopyran-4-one (Figure 1).¹² Hesp is

produced from the alkaline hydrolysis of hesperetic acid and phloroglucinol, which during acid hydrolysis gets converted into hesperetin, l-rhamnose, and d-glucose.^{12,16} The biological activity of Hesp is due to the presence of hydroxyl moieties in both the heterocyclic and aromatic rings.^{12,16} Further, there is a close relationship between the presence and number of hydroxyl moieties with the potential antioxidant capacity of hesperidin.^{12,16,17} Additionally, Hesp also harbors neuro-protective actions and has the capability of penetrating the blood-brain barrier.^{12,16,17}

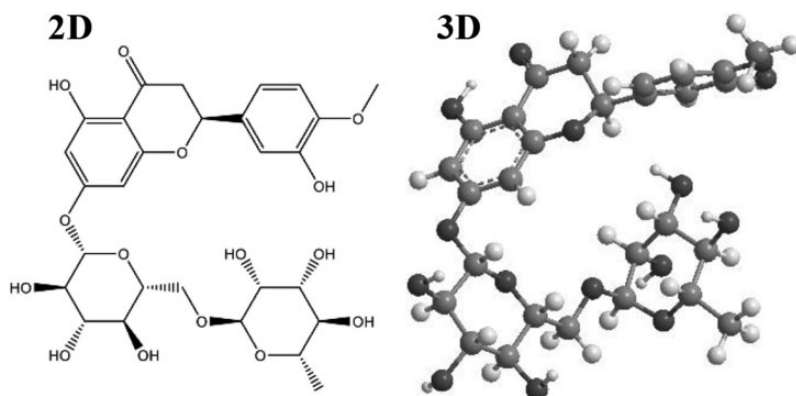
Hesperidin: Molecular mechanisms of action

Apoptotic and cell cycle arrest

Apoptosis induction and cell cycle arrest are among the most important mechanisms of Hesp action against cancer cells. Studies revealed that the pro-apoptotic action of Hesp is related to different kinase pathways. For example, suppression of phosphatidylinositol-4,5-bisphosphate 3-kinase subunit/ AKT Serine/threonine kinase/ inhibitor of kappa light polypeptide gene enhancer in B-cells (PI3K/ Akt/IKK) signalling pathway in NALM-6 human pre-B cells by Hesp was reported to induce apoptosis (Figure 2).¹⁸ In breast cancer cell line, MCF-7, Hesp induced apoptotic events like phosphatidyl-serine externalization, DNA fragmentation, caspase-7 activation, and PARP (Poly (ADP-ribose)polymerase) cleavage, which are associated with the activation of caspase-9, loss of mitochondrial membrane potential, release of cytochrome c, and an increase Bax:Bcl-2 ratio. Further experiments revealed that Hesp induced apoptosis by accumulating reactive oxygen species (ROS) and activation of apoptosis signal regulating kinase 1/ Jun N-terminal kinase (ASK1/JNK) pathway.¹⁹ Besides these, the most important mechanism of apoptotic effect of Hesp is via generation of ROS. As reported by Zhang *et al.*,²⁰ high ROS levels, along with adenosine triphosphate (ATP) and calcium are responsible for the induction of apoptosis by Hesp in hepatocellular carcinoma cells via the activation of mitochondrial pathway. Similarly, in gastric cancer cells and esophageal cancer cells, Hesp induces apoptosis by increasing ROS and activating mitochondrial pathway.^{21,22}

Caspase-dependent apoptosis induction by Hesp in NCI-H358 and A549 non-small cell lung cancer (NSCLC) cells was also observed by Birsu *et al.*²³ and Xia *et al.*²⁴, while Hesp was also found to arrest cell cycle at G₀/G₁ phase via the downregulation of cyclinD1, and increasing the expression of p21 and p53. In addition, the endoplasmic reticulum stress pathway was also found to be involved in apoptosis induction in HeLa (immortal cervical cancer cells) by Hesp, along with arresting cell cycle at G₀/G₁ phase via the downregulation of cyclin D1, cyclin E1, and cyclin-dependent kinase 2 (Cdk2) at protein level²⁵. The ROS-mediated apoptosis induction by hesp in human gall bladder carcinoma was also reported by Pandey *et al.*,²⁶ but they found cell cycle arrest at G₂/M phase. A recent²⁵ evidence showed that Hesp is a potent inhibitor of calcium/calmodulin-dependent protein kinase IV

Hesperidin



Hesperetin

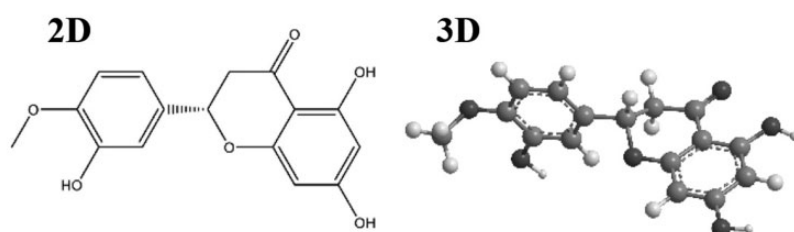


Figure 1. The chemical structures and three-dimensional (3D) conformers of Hesperidin and Hesperetin.

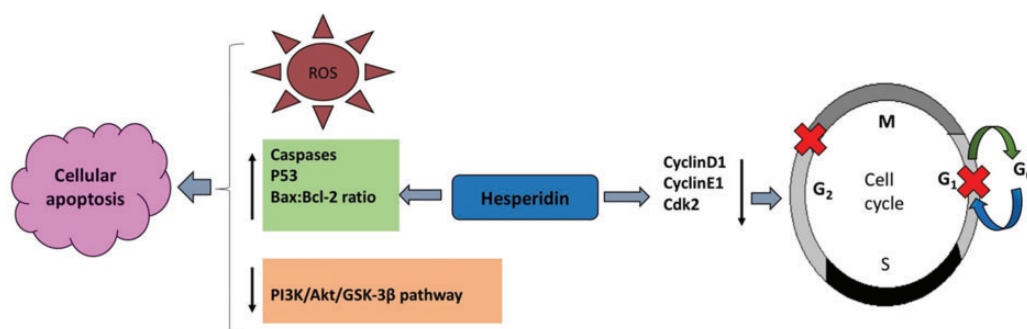


Figure 2. Role of hesperidin on apoptosis and cell cycle. Hesperidin can generate reactive oxygen species (ROS) in cancer cells and activate mitochondrial pathways (by upregulating caspases) and inhibit kinases, which can induce apoptosis. Also by regulating cell cycle-related proteins, hesperidin can arrest cancer cell cycle in G₀/G₁ phase and G₂/M phase. (A color version of this figure is available in the online journal.)

(CAMKIV), and by inhibiting CAMKIV along with activating the caspase-3-dependent intrinsic pathway through the upregulation of pro-apoptotic protein, Bax (BCL2 associated X, apoptosis regulator), Hesp exerts its anti-apoptotic and anticancer activities.²⁷

The *in vitro* activity of Hesp in inducing apoptosis and arresting cell cycle is also proved *in vivo* (Table 1). For example, in azoxymethane-induced mouse model of colon cancer, Hesp was found to alter the anti-apoptotic scenario by modulating Bax/Bcl-2 ratio, together with enhanced release of cytochrome-c and activation of caspase-3/9. Experimental studies revealed that Hesp initiates apoptosis by inhibiting constitutively activated Aurora-A-mediated PI3K/Akt/GSK-3 β pathway, and mTOR (mammalian

target of rapamycin) pathway coupled with the stimulation of autophagy in this colon cancer model.²⁸ Hesp in combination with fistein has been reported to inhibit cellular proliferation via triggering programmed cell death in human K562 chronic myeloid leukemia (CML) cells through activation of caspase-3 and JAK/STAT (Janus Kinase/Signal transducer and activator of transcription 3) pathway and genes of JAK/STAT pathway have also been identified as candidates of CML therapy.²⁹ In ferric nitrilotriacetate (Fe-NTA)-induced renal cancer model of Wistar rats, Hesp was found to induce apoptosis-related proteins caspase-3, caspase-9, Bax expression and downregulation of Bcl-2 (BCL2 apoptosis regulator), NF- κ B (nuclear factor kappa B subunit), iNOS (inducible nitric oxide synthase), TNF- α

Table 1. A brief overview of the *in vivo* studies carried out using hesperidin.

Compound of interest	Model of study	Type of cancer	Treatment dose	Underlying mechanisms of observed results	Refs
Hesperidin	Diethylnitrosamine/ CCI4-induced rats	Hepatocellular carcinoma	11 mg/kg	↓oxidative stress, inflammation, cell proliferation, TGF- β 1/Smad3 signaling, and collagen deposition by activating Nrf2/ARE/HO-1 and PPAR γ pathways	32
	Diethyl nitrosamine hepatocarcinogenesis-induced rat	Hepatocellular carcinoma	1000, 500, and 250 ppm	Hypomethylating effect on the LINE-1 sequence (up to 47% hypomethylation at 12.5 mM) and on the ALU-M2 repetitive sequences (up to 32% at 6 mM)	33
	Rats	Hepatocellular carcinoma	150 mg/kg/day	↓Wnt3a, β -catenin, Cyclin D1, and Wnt5a gene expressions	34
	Female Sprague-Dawley rats	Breast cancer	30 mg/kg/body weight	Elevation in glycoproteins, nucleic acids, lysosomal enzymes and also significant alterations in macromolecules in renal tissues of cancer bearing animals	35
	Rats	Hepatocellular carcinoma	50, 100, and 200 mg/kg/d	↓Exosomal RAB11A messenger RNA and long noncoding RNA-RP11-583F2.2 along with the increase in exosomal miR-1298	36
	Wistar rats	Renal cancer	100 and 200 mg/kg body weight	↓PGE2, COX-2, VEGF, and improved renal function by ↓BUN, creatinine, and KIM-1	37
	Wistar rats	Renal cancer	100 and 200 mg/kg b wt	Induce caspase-3, caspase-9, bax expression, and downregulate bcl-2, NF κ B, iNOS, TNF- α , PCNA expression	30
	Wistar rats	Hepatocellular carcinoma	200 mg/kg body weight	↓PI3K, Akt, CDK-2 protein expression	38
	Mice	Lung cancer	25 mg/kg body weight	↓COX-2, MMP-2, and MMP-9	39
	Colon carcinoma (CT-26)-bearing mice	Colon cancer	200 mg/kg	↓WBC count caused by cyclophosphamide to reduce antitumor effect	40
	7, 12-Dimethylbenz (a) anthracene-induced rats	Breast cancer	30 mg/kg	Decline in lipid peroxidation and membrane bound marker enzyme AST, ALT, ALP, ACP, 5'ND, γ -GT	41
	Benzo(a)pyrene-induced Swiss albino mice	Lung cancer	50 mg/kg	↓lipid peroxides, aryl hydrocarbon hydroxylase (AHH), gamma glutamyl transpeptidase (γ -GT), 5'-nucleotidase (5'-ND) and lactate dehydrogenase (LDH)	42

(tumor necrosis factor-alpha), PCNA (proliferating cell nuclear antigen) expression, which were associated with Hesp anti-cancer activities *in vivo*.³⁰ In xenograft model of colon cancerous mice, Hesp also showed mitochondrial pathway-mediated apoptosis induction, and cell cycle arrest at G₂/M phase.³¹

Anti-angiogenesis and anti-metastasis

In view of the limitations of presently marketed monoclonal antibodies and synthetic compounds as anti-angiogenic agents pertaining to toxicity and high cost incurred, natural compounds are being extensively explored as potential anti-angiogenic agents.^{43,44} These natural products such as flavonoids (e.g. hesp) modulate tumor angiogenesis via targeting vascular endothelial growth factor (VEGF), matrix metalloproteinases (MMPs), basic fibroblast growth factor (bFGF), endothelial cell proliferation, migration, and metastasis owing to their anti-proliferative potential.⁵ The first excerpts of the anti-metastatic and anti-proliferative effect of hesperetin were documented in 2007 by Lentini *et al.*⁴⁵ in B16-F10 metastatic murine melanoma cells *in vitro* and C57BL6/N mice *in vivo*. Thereafter,

in 2009, Yeh *et al.*⁴⁶ illustrated anti-metastatic potential of Hesp *in vitro* in HepG2 human hepatocellular carcinoma cells where hesperidin suppressed secreted cytosolic MMP-9 expression via inhibition of activator protein-1 (AP-1, JNK signaling pathway) and NF- κ B (NF- κ B signaling pathway). This was further supported by findings from other studies in 2010 which reported Hesp inhibited 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced cytosolic MMP-2 and MMP-9 and cyclooxygenase-2 (COX-2) expression via modulating NF- κ B and AP-1 induced tumor cell invasion and metastasis in lung cancer and hepatocellular carcinoma (Figure 3).^{39,47} Later, in 2012, hesperetin was documented to inhibit TGF- β 1 (transforming growth factor- β signaling pathway)-induced tumor migration and metastasis via phosphorylation of Smad3 (SMAD family member 3).⁴⁸

In 2015, Kim⁴⁹ reported the inhibitory effect of Hesp on vascular formation in human umbilical vascular endothelial cells (HUVECS) and mouse embryonic stem cell (mES)-derived endothelial like cells via blocking AKT and mTOR signaling pathway which lead to inhibition of cell migration, suppression of micro-vessel sprouting, and tube

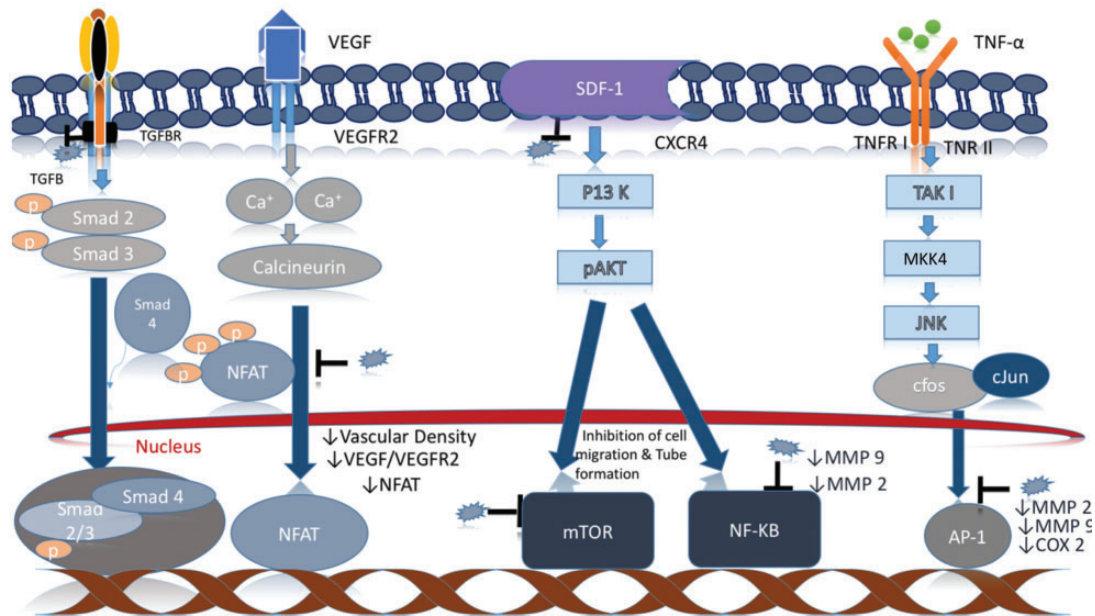


Figure 3. Cellular targets of hesperidin while acting as anti-angiogenic and anti-metastatic agent. (A color version of this figure is available in the online journal.)

formation in HUVECS. Zhao *et al.*⁵⁰ documented the anti-angiogenic effect of Hesp (a component of Qingdu granule) in MCF-7 and HUVECs cells where Hesp inhibited migration and tube formation in human breast cancer cells via downregulation of NFATc3 (nuclear factor of activated T-cells) expression. In female BALB/c nude mice, a xenograft tumor model *in vivo* Hesp inhibited tumor growth, decreased vascular density, inhibited VEGF, and downregulated NFATc3, VEGF, and VEGFR2 expression via NFAT signaling pathway.⁵⁰ Further research in the domain of anti-metastatic potential of Hesp in pancreatic cancer has shown to target MKK3/6 and p38 intracellular signaling pathways.⁵¹ In A549 non-small cell lung cancer cells, Hesp was described to significantly inhibit tumor migration capability via targeting SDF-1 α (stromal cell-derived factor 1) leading to downregulation of CXCR-4 (C-X-C chemokine receptor type 4), p-Akt, p-I κ B (phosphorylated-I kappa B), and p-p65 expression (SDF-1/CXCR-4 signaling cascade).⁵² Hesp was also documented to inhibit cell migration and invasion in human osteosarcoma MG-63 cells via wound healing and matrigel assay and *in vivo* in male BALB/c xenograft mice model.³¹

Hesperetin administered in combination with platinum drugs *in vitro* in A549 lung adenocarcinoma cells and *in vivo* in C57BL/6 mice inhibited tumor proliferation and migration via targeting UDP-glucuronosyltransferase (UGT) family 1 member A3 (UGT1A3) more significantly in comparison to single drug treatment regime⁵³. In another study, combined administration of naringenin with hesperetin was shown to maximize anti-metastatic effect in Panc-1 human pancreatic cancer cells via downregulation of FAK (focal adhesion kinase) and p38 signaling pathway⁵⁴. These *in vitro* (Table 2) and *in vivo* studies (Table 1) on the cumulative front document the potential anti-angiogenic and anti-metastatic potential of hesperidin which may be used as a promising anti-cancer strategy for the management of

human cancers without eliciting toxic effects on surrounding normal cells.

Anti-oxidant and anti-inflammatory effects

Inflammation is a complex physiological and biological process in which body fights toward the harmful stimuli such as undigested particles, chemical irritants, damaged cells, moreover viral, bacterial and parasitic infections by overexpression of various cytokines, chemokines, and pro-inflammatory mediators, including TNF- α , COX-2, IL-1 β (interleukin-1 β), IL-6, IL-8, iNOS, NO (nitric oxide), prostaglandins, and eicosanoids.⁶⁰⁻⁶⁵ Apart from the vigorous dependence of the expression level of pro-inflammatory mediators on the activation of different signaling pathways that can be regulated by various factors such as MAPKs (mitogen-activated protein kinases), NF- κ B, ICAM-1 (intercellular adhesion molecule-1), and VCAM-1 (vascular cell adhesion molecule-1), there is a tight relationship between inflammation and production of ROS and RNS (reactive nitrogen species).⁶⁶⁻⁷¹ Moreover, the mounting evidences indicate that both inflammation and oxidative stress drive carcinogenesis by inactivation of tumor suppressor genes, activation of oncogenes, and disruption of various cellular signaling pathways.^{72,73} On the other hand, it is a wide thought that Hesp and its derivatives can be considered as the substantial and effective traditional flavonoids on oxidative stress and inflammation along with proliferation, apoptosis, DNA damage, free radicals, carcinogenesis, hypertension, hyperglycemia, and hypolipidemia.^{74,75} The studies on anti-inflammatory mechanisms of Hesp indicate that Hesp can reduce the level of inflammatory factors such as VCAM-1, COX-2, MMP-2, MMP-9, PGE2 (prostaglandin E2), IL-4, IL-6, iNOS, and NO₂.^{74,76} The anti-oxidative, chelating, and strong reducing properties along with the hydroxyl, hydrogen peroxide, superoxide, and free radical scavenging activities of Hesp occurred depending on the

Table 2. A brief overview of the *in vitro* studies carried out using hesperidin to study effect on cellular processes.

Effect	Underlying mechanism	Concentration	Cell line	Cancer type	Refs
Apoptosis	↑Caspase-9, -8, and -3 activities, Bax, Bak, and tBid protein levels ↓Bcl-xL protein	150.43 ± 12.32 μM	HepG2	Hepatic cancer	55
	↑caspase-3 and ↓mitochondrial membrane potential	50 μM	A549 and NCI-H358	Lung cancer	23
	cleavages of Bid, caspase-3, and PARP, upregulation of Bax, and down-regulation of Bcl-xl	152.3 μM	MSTO-211H	Human malignant pleural mesothelioma	56
	↑caspase-3 cleavage expression through ADD153'CHOP'GRP78 and cytochrome c signaling pathways	10 μM	A2780	Ovarian Cancer	57
	↑expression of PPARγ and p53 accumulation, ↓NF-κB	10 μM	NALM-6	Lymphoblastic leukemia	58
	↑ROS, nuclear condensation, and activation of Caspase-3	143.39 mM	Primary GBC	Gall bladder carcinoma	26
Cell cycle arrest	G2/M phase cell cycle arrest	33.5, 23.8 and 17.6 μM, respectively, at 24, 48 and 72 h	MG-63	Cervical cancer	31
	G0/G1 arrest by ↓cyclinD1, cyclinE1, and cyclin-dependent kinase 2 at the protein level.	0, 40, 80, and 160 μM	HeLa, HT-29	Cervical cancer and Colon cancer	
Anti-metastatic	G0/G1 arrest by ↓cyclin D1 and ↑p21, p53	75–125 μg/mL	A549	Lung cancer	24
	↓acetaldehyde-activated NF-κB and activator protein 1 (AP-1) activity results in ↓MMP-9 expression	50 μM	HepG2	Hepatocellular carcinoma	46
	↓MMP-9 via NF-kappaB an AP-1 signaling pathway	50 μM	HepG2	Hepatocellular carcinoma	47
	14-fold increase in loss of MMP via FGF and NF-κB signal transduction pathways	50 μM	A549	Lung cancer	23
	inhibit migratory and invasive capability by SDF-1/CXCR-4 signaling cascade	25–65.5 μg/mL	A549	Lung cancer	52
Anti-angiogenic	inhibits vascular formation by blocking the AKT/mTOR signaling pathways	100 μM	HUVECs	Nil	49
Anti-oxidant	Cytotoxicity mediated through antioxidant properties	IC50 recorded at b10 μg/mL	MCF-7, Hep-2, HeLa and HepG-2	Breast, larynx, cervix and liver carcinoma	59

concentration and originated from the chemical structure by acting as a hydrogen donor to the radical molecules, and played a radical target role to form new complexes between the antioxidant radicals and the lipid radicals thanks to the presence of 300-hydroxy, 400-o-methoxy system in the B ring.²

Moreover, the radical scavenging activity is increased due to the entity of 300-OH and 5-OH groups, in combination with a 4-carbonyl function and C40–C80 double bond though the anti-oxidant activity of Hesp can be considered as moderate according to the other flavonoids due to the lack of OH at C4.^{2,9,68,77} On the other hand, the non-glycosylated form of Hesp named hesperetin displays a stronger anti-oxidant activity than hesp because it has an additional hydroxyl group in its molecular structure and O-glycosylation reduces the antioxidant activity along with the electronic delocalization capacity (Figure 1).^{12,78} The anti-oxidant activity of Hesp and hesperetin, which can be achieved by boosting cellular anti-oxidant defense and radical scavenging, has been verified by various *in vivo* and *in vitro* experimental models.^{2,74} For example, it was reported that Hesp could protect the cellular components such as DNA and proteins by radical scavenging and ROS neutralizing activity.^{12,76,79} Moreover, it has been proved in

the experimental models that Hesp counteracts the harmful activities of hydrogen peroxide, peroxyxynitrite, carbon tetrachloride, cadmium, acetaminophen, nicotine, cyclophosphamide, acrylonitrile, dimethylbenz[a]anthracene, tert-butyl hydroperoxide, lipopolysaccharide, benzo[α]pyrene, technetium, gamma radiation and many others by increasing the level and activity of antioxidant enzymes and compounds such as GSH (reduced glutathione), GPx (glutathione peroxidase), GST (glutathione S-transferase), GR (glutathione reductase), SOD (superoxide dismutase), CAT (catalase), vitamin C and vitamin A, and decreasing the level of cellular damage markers such as LDH (lactate dehydrogenase), LPO (lipid peroxides), GGT (gamma glutamyl transpeptidase), AHH (aryl hydrocarbon hydroxylase), 5 ND (5 nucleotidase), ALP (alkaline phosphatase), AST (aspartate aminotransferase), ALT (alanine aminotransferase), and bilirubin.^{76,80,81}

The Hesp and hesperetin induced increase in the level and activity of the anti-oxidant enzymes can be achieved via Keap1-Nrf2 (nuclear factor erythroid 2-related factor 2) pathway, which is known as the major regulator of oxidative and electrophilic stress.⁷⁵ Briefly, Hesp and hesperetin increase the expression of Nrf2, separate Keap1-Nrf2 complex, and increase the nuclear translocation of Nrf2, and the

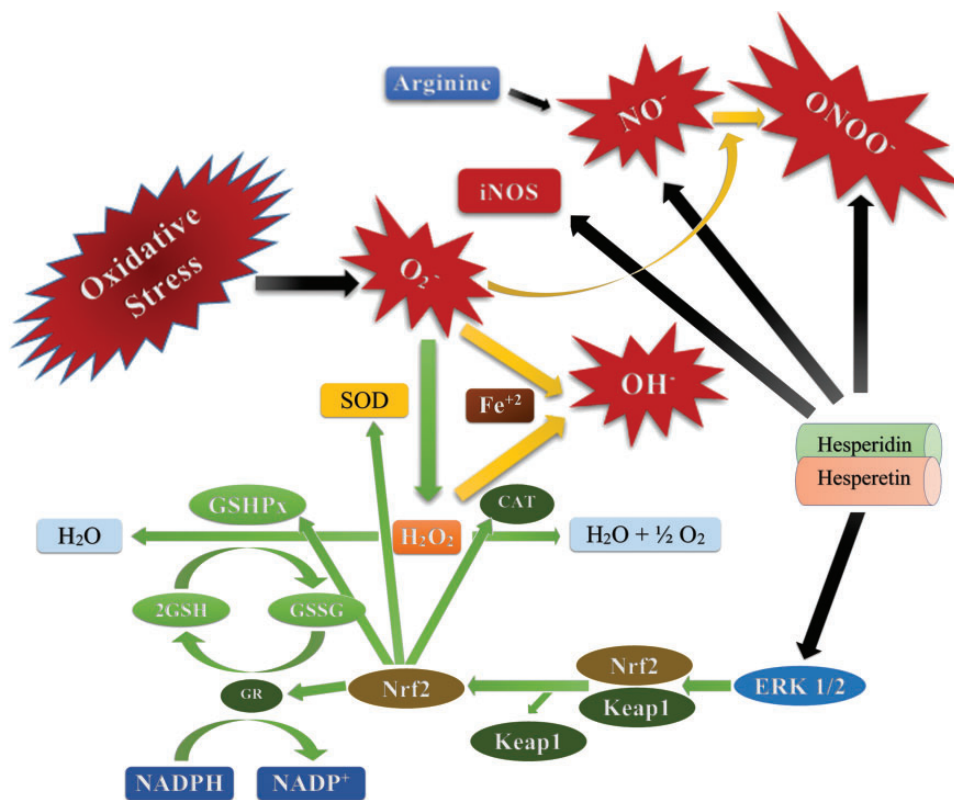


Figure 4. The anti-oxidant activity mechanisms of hesperidin. (A color version of this figure is available in the online journal.)

production of anti-oxidant enzymes is increased by the activation of gene transcription thanks to the binding of Nrf2 to the antioxidant response element (ARE) within gene promoter region.^{75,82} Further, Hesperetin derivative-14 (HD-14) has also been reported to have anti-inflammatory potential and has been shown to inhibit p-JAK1/p-STAT2 via PPAR- γ upregulation in LPS-treated RAW264.7 cells.⁸³ Similar results were reported using another Hesperetin derivative-12 (HDND-12) in RAW264.7 cells and were reported to down-regulate p-JAK2/p-STAT3 expression.⁸⁴ Consequently, the derived perception from the published studies about hesperidin and its derivatives indicates that the anticancer property of Hesp seems to be largely originated from its anti-oxidant and anti-inflammatory activity (Figure 4).

Synergistic effect of hesperidin with other anti-cancer agents

Hesp, a flavanone, is present in different citrus fruits and possesses a number of biological activities.⁸⁵ Hesp is identified to possess potent anti-inflammatory, anti-carcinogenic, and anti-oxidant activities in different studies.⁵ The chemical structure of Hesp consists of hesperetin (methyl eriodictyol) bound to rutinose.¹² The glycoside entity of Hesp is a disaccharide that comprises rhamnose and glucose.¹² The research in present times on anticancer activities of natural compounds has been targeted on induction of cancer cell death. These natural compound induces

cancer cell death initiated by apoptosis (type I programmed cell death), autophagic cell death (type II programmed cell death), and necroptosis (programmed necrosis).⁸⁵⁻⁸⁸ Hesp and Hesperetin cause cell proliferation delay in different cancer models. Different effects of these compounds have been reported depending on the factors including dose, type of compound, and cell line under study (Table 3). Cell cycle arrest related to cytostatic effects has been reported in cells that have elevated p53 and cyclin-dependent kinase inhibitor levels, along with lowered levels of cyclins and cyclin-dependent kinases.⁵⁵

Synergistic interaction of hesperidin with different compounds

Doxorubicin

Doxorubicin (DOX) is widely employed in anti-tumour therapies.^{94,95} In spite of the systematic chemotherapy with doxorubicin, it provides only peripheral improvements in a survival of the hepatocellular carcinoma patients.^{96,97} The main mode of action of doxorubicin includes intercalation within DNA base pairs, thus resulting in DNA strand breakage and an inhibition of DNA and RNA synthesis by inhibition of topoisomerase II, resulting in DNA damage and apoptosis induction.⁹⁸ The doxorubicin also initiates the ROS generation thus causing cell death. The application of apigenin and Hesp alongside doxorubicin revealed the effect on doxorubicin-induced

Table 3. Synergistic relationship of hesperidin with other natural compounds against cancer.

Molecules in synergism	Effects	Refs
Doxorubicin – hesperidin	Alterations in the expression levels of HK2 and LDHA	89
Doxorubicin – hesperidin	PgP expression inhibition	90
Cytarabine – hesperidin	Lowering in IC50 values of Cytarabine	91
Tamoxifen – hesperidin	Apoptosis induction, cell cycle arrest and downregulation of EGFR (epidermal growth factor receptor) and ER α (estrogen receptor alpha).	92
Quercetin – hesperidin	H ₂ O ₂ scavenging	99

toxicity. These compounds altered the expression levels of glycolytic pathway genes – HK2 (hexokinase 2) and LDHA (lactate dehydrogenase A), which possess a major role in the Warburg effect. The simultaneous administration of doxorubicin and apigenin or Hesp eradicated the damage with the simultaneous increase in the doxorubicin toxicity. In another study, effect of Hesp has been investigated in combination with doxorubicin to check its effect on doxorubicin-resistant MCF7 breast cancer cell lines. The cytotoxic effects were studied using MTT assay. Hesp combined with doxorubicin was unable to increase the apoptotic initiation but inhibited the PgP (P-glycoprotein) expression⁹⁰ which is mainly responsible for developing the multidrug resistance after administration of doxorubicin.

Cytarabine

In another study, the use of Silibinin and Hesp showed 50% cell inhibition at 16.2 μ M and 50.12 μ M, respectively,⁹¹ whereas the fixed doses of Hesp and Silibinin in conjunction with Cytarabine at different concentrations lowered the IC 50 value of Cytarabine by 5.9 and 4.5 folds, respectively. Drug interaction analysis exhibited that Silibinin and Hesp showed synergistic effect with Cytarabine in 1:50 to 1:250 ratios.⁹¹ Hence, these compounds can be employed as chemotherapeutic agents either alone or in combination.

Tamoxifen

Tamoxifen (Tam) is a commonly used anticancer drug for estrogen receptor (ER)-positive breast cancer treatment. In different studies, it was shown that the some natural compounds, like Hesp, piperine (Pip) and bee venom (BV) have inhibitory effect on the breast cancer cells growth when used individually. The combined effect of these natural compounds and Tam was investigated in a study with a hypothesis that these compounds can increase the potential efficacy of growth inhibitory activity of Tam. The cytotoxic activity of Hesp, BV, and Pip was examined on MCF7 and T47D breast cancer cell lines via MTT assay and achieved equitable IC50 comparable to Tam results.⁹² The effect of different combinations was investigated and enhanced anti-proliferative result was obtained on MCF7 and T47D cell lines due to the synergistic effect. These natural compounds can synergistically increase the potential anticancer property of Tam against MCF7 and T47D cells probably by inducing the apoptosis, cell cycle size and EGFR (epidermal

growth factor receptor), and ER α (estrogen receptor alpha) downregulation.

Quercetin

Etoposide [40-demethylepipodo- phyllotoxin- 9-(4,6-O-ethylidene) -b-d glucopyranoside] is a derivative of natural product, podophyllotoxin. Etoposide acts as therapeutic agent in different types of cancer due to its ability to inhibit the topoisomerase II enzyme and induction of DNA breaks. In a study, the synergistic effect of Hesp and quercetin was studied to improve oxidative damage caused by Etoposide on reproductive system in male rats. In this study, it was observed that administration of quercetin at concentration levels of 20 mg/kg body weight and Hesp at 25 mg/kg body weight for two months significantly enhanced sperm motility and count in experimental groups in comparison to etoposide-treated group. This improvement in sperm motility and count can be attributed to the H₂O₂ scavenging by quercetin and Hesp, resulting in inhibition of cellular DNA damage^{93,99}.

Role of nano-technology in hesperidin delivery

Natural anti-oxidants such as hesperidin has shown promising impact for the treatment of malignant growth and other alike diseases due to high efficacy and lower side effects as compared to synthetic drugs.^{100,101} But Hesp's clinical use was extremely restricted due to lower aqueous solubility and poor bioavailability.¹⁰² So there is need to overcome these issues for optimal use of this compound.¹⁰³ Nanotechnology is an interdisciplinary area of research having broad applications like molecular imaging, molecular diagnosis, and particularly targeted drug delivery.¹⁰⁴ Further, this technology can also overcome the solubility and bioavailability issues of drugs¹⁰⁵ as these parameters have tremendous impact on treatment of cancer.¹⁰⁶ Therefore, several studies were initiated on developing Hesp-based nanoparticles to improve the bioavailability, absorption, and bio-distribution of this flavonoids.^{103,106,2525-109} Gu *et al.*¹⁰⁶ have developed two nano-based formulations, i.e. hesperetin-TPGS (D- α -tocopheryl polyethylene glycol 1000 succinate) micelles and hesperetin-phosphatidylcholine (PC) complexes to improve the water solubility, antioxidant activity, and oral absorption of hesperetin and these formulation led to the increase of 16.2- and 18.0-fold in *in-vitro* antioxidant potential and *in vivo* oral absorption of hesperetin.

Similarly, Duranolu *et al.*¹⁰³ developed hesperetin-loaded nanoparticles with high encapsulation efficiency and low particle size and various process parameters were optimized for the optimal use of hesperetin. Further to develop new targeting strategies Ferrari *et al.*¹⁰⁸ have developed Hesp-coated solid lipid nanoparticles and evaluated their various physicochemical properties for optimum-targeted delivery for the treatment of several disorders. Praveen Kumar *et al.*¹⁰⁹ evaluated cerebroprotective potential of hesperidin nanoparticles for the effective treatment of cerebral ischemia in rats. To increase the drug delivery potential for topical applications, Menezes *et al.*¹¹⁰ reported to fabricate textile-based Hesp-loaded nanocapsules. These fabric-based nanocapsules were found to be suitable for sustained release of drug. Although efforts were initiated for the fabrication of Hesp-coated nanoparticles for the effective treatment of various disorders, but there is lacunae of reports where Hesp-coated nanoparticles were clinically tried for cancer treatment. Therefore, more progressive endeavors are required to integrate hesperidin-based nanoparticles for the effective treatment of cancer and other diseases.

Conclusion and future perspectives

The importance of citrus bioflavonoids is attributed to the pharmacological activity of Hesp. Despite extensive treatment protocols and regimes for cancer patients ranging from surgery to chemotherapy to immunotherapy, cancer is not completely curable and the conventional treatment regime is associated with short-term and long-term adverse events. Recently, lot of research is ongoing to explore bioflavonoids, e.g. Hesp in cancer treatment in view of their potential antioxidant property. Recent literature from different research groups have emphasized on the potent antioxidant capacity of Hesp and its potential role as anticancer agent. This review demonstrates the anti-tumor effects of Hesp in different malignancies with special emphasis on its molecular mechanism of action. The importance of Hesp as anti-cancer agents is evident from the experimental evidences emphasizing on how Hesp modulates oxidative stress, inflammation, and cancer cell death (hallmarks of cancer). Further, Hesp has been reported to augment apoptosis in malignant cells via NF- κ B, mTOR, PI3K/AKT pathways.¹¹¹ Hesp has also been documented to down-regulate pro-inflammatory mediators and enzymes (IL-1/6, TNF, COX-2) in tumorigenesis and improve anti-oxidant defense mechanism. In addition, Hesp significantly improves pharmacological symptoms of other diseases such as arthritis, myocardial, and infertility.¹¹²⁻¹¹⁶ Therefore, main benefit of using flavonoids in contrast to chemotherapeutic agents is attributed to their low toxicity and tolerability, and it is worthwhile to mention that even at highest dose, Hesp/hesperetin do not cause cytotoxic effects or acute oxidative damage. However, further studies are required to unravel the therapeutic effects of Hesp/hesperetin in cancer treatment. Though, Hesp is presently in pre-clinical trials, promising data from clinical trials are warranted to increase the translation applicability of Hesp in cancer treatment. From the future perspective, future

in vitro and *in vivo* studies focusing on the following dimensions of Hesp need to be studied to translate the practical applicability of Hesp as anticancer agent:

1. To increase the bioavailability and absorption of Hesp/hesperetin (aglycone form).
2. Defining the precise molecular mechanisms of action of hesp through meticulously designed and executed experimental studies for anticancer effects Hesp.
3. Standardizing the optimum effective dose of Hesp for translation in clinical trials in combination with conventional chemotherapeutic agents and targeted therapies.
4. Evaluation of safety and efficacy findings in cancer patients undergoing treatment with Hesp.

Authors' contributions: VA and HST: performed literature survey, data extraction and compiled the manuscript; KS: wrote the introduction section; Md. AK: Contributed in role of hesperidin in apoptosis induction and cell cycle arrest; VA composed the text on chemistry of hesperidin and its role in modulating angiogenesis and metastasis; SS and AP: prepared the section synergistic interaction of hesperidin with different compounds; MV; Composed the text of anti-oxidant and anti-inflammatory activity of hesperidin; DA wrote role of nanotechnology in hesperidin delivery; FT and PS: helped with conclusion sections, design and drew figures of the indicated sections and prepared the *in vivo* and *in vitro* study tables for the hesperidin manuscript; GS: critically reviewed and edited the manuscript; All authors read and approved the final manuscript.

ACKNOWLEDGMENTS

The authors acknowledge the Post Graduate Institute of Medical Education and Research (PGIMER), Chandigarh and Maharishi Markandeshwar (Deemed to be University), Mullana-Ambala for providing an opportunity to complete the review.

DECLARATION OF CONFLICTING INTERESTS

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

FUNDING

The author(s) received no financial support for the research, authorship, and/or publication of this article.

ORCID iD

Gautam Sethi  <https://orcid.org/0000-0002-8677-8475>

REFERENCES

1. Barreca D, Gattuso G, Bellocco E, Calderaro A, Trombetta D, Smeriglio A, Lagana G, Daglia M, Meneghini S, Nabavi SM. Flavanones: citrus phytochemical with health-promoting properties. *Biofactors* 2017;43:495-506

2. Ahmadi A, Shadboorestan A. Oxidative stress and cancer; the role of hesperidin, a citrus natural bioflavonoid, as a cancer chemoprotective agent. *Nutr Cancer* 2016;**68**:29–39
3. Li C, Schluesener H. Health-promoting effects of the citrus flavanone hesperidin. *Crit Rev Food Sci Nutr* 2017;**57**:613–31
4. Aishatwi AA, Ramesh E, Periasamy VS, Subash-Babu P. The apoptotic effect of hesperetin on human cervical cancer cells is mediated through cell cycle arrest, death receptor, and mitochondrial pathways. *Fundam Clin Pharmacol* 2013;**27**:581–92
5. Roohbakhsh A, Parhiz H, Soltani F, Rezaee R, Iranshahi M. Molecular mechanisms behind the biological effects of hesperidin and hesperetin for the prevention of cancer and cardiovascular diseases. *Life Sci* 2015;**124**:64–74
6. Ahmadi A, Shadboorestan A, Nabavi SF, Setzer WN, Nabavi SM. The role of hesperidin in cell signal transduction pathway for the prevention or treatment of cancer. *Curr Med Chem* 2015;**22**:3462–71
7. Nagai H, Kim YH. Cancer prevention from the perspective of global cancer burden patterns. *J Thorac Dis* 2017;**9**:448–51
8. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018;**68**:394–424
9. Wang CZ, Zhang Z, Anderson S, Yuan CS. Natural products and chemotherapeutic agents on cancer: prevention vs. treatment. *Am J Chin Med* 2014;**42**:1555–8
10. Bishayee G, Sethi G. Bioactive natural products in cancer prevention and therapy: progress and promise. *Semin Cancer Biol* 2016;**40–41**:1–3
11. Dai X, Zhang J, Arfuso F, Chinnathambi A, Zayed ME, Alharbi SA, Kumar AP, Ahn KS, Sethi G. Targeting TNF-related apoptosis-inducing ligand (TRAIL) receptor by natural products as a potential therapeutic approach for cancer therapy. *Exp Biol Med* 2015;**240**:760–73
12. Garg A, Garg S, Zaneveld LJ, Singla AK. Chemistry and pharmacology of the citrus bioflavonoid hesperidin. *Phytother Res* 2001;**15**:655–69
13. Mesquita E, Monteiro M. Simultaneous HPLC determination of flavonoids and phenolic acids profile in Pera-Rio orange juice. *Food Res Int* 2018;**106**:54–63
14. Shimoda K, Hamada H, Hamada H. Glycosylation of hesperetin by plant cell cultures. *Phytochemistry* 2008;**69**:1135–40
15. Perfetti GA, Joe FL, Jr., Fazio T, Page SW. Liquid chromatographic methodology for the characterization of orange juice. *J Assoc off Anal Chem* 1988;**71**:469–73
16. Ranganna S, Govindarajan VS, Ramana KV. Citrus fruits - varieties, chemistry, technology, and quality evaluation. Part II. Chemistry, technology, and quality evaluation. A. chemistry. *Crit Rev Food Sci Nutr* 1983;**18**:313–86
17. Mitsunaga Y, Takanaga H, Matsuo H, Naito M, Tsuruo T, Ohtani H, Sawada Y. Effect of bioflavonoids on vincristine transport across blood-brain barrier. *Eur J Pharmacol* 2000;**395**:193–201
18. Shahbazi R, Cheraghpour M, Homayounfar R, Nazari M, Nasrollahzadeh J, Davoodi SH. Hesperidin inhibits insulin-induced phosphoinositide 3-kinase/akt activation in human pre-B cell line NALM-6. *J Cancer Res Ther* 2018;**14**:503–8
19. Palit S, Kar S, Sharma G, Das PK. Hesperetin induces apoptosis in breast carcinoma by triggering accumulation of ROS and activation of ASK1/JNK pathway. *J Cell Physiol* 2015;**230**:1729–39
20. Zhang J, Song J, Wu D, Wang J, Dong W. Hesperetin Induces the Apoptosis of Hepatocellular Carcinoma Cells via Mitochondrial Pathway Mediated by the Increased Intracellular Reactive Oxygen Species, ATP AND Calcium. *Med Oncol* 2015;**32**:101
21. Wu D, Zhang J, Wang J, Li J, Liao F, Dong W. Hesperetin induces apoptosis of esophageal cancer cells via mitochondrial pathway mediated by the increased intracellular reactive oxygen species. *Tumour Biol* 2016;**37**:3451–9
22. Zhang J, Wu D, Vikash Song J, Wang J, Yi J, Dong W. Hesperetin induces the apoptosis of gastric cancer cells via activating mitochondrial pathway by increasing reactive oxygen species. *Dig Dis Sci* 2015;**60**:2985–95
23. Birsu Cincin Z, Unlu M, Kiran B, Sinem Bireller E, Baran Y, Cakmakoglu B. Anti-proliferative, apoptotic and signal transduction effects of hesperidin in non-small cell lung cancer cells. *Cell Oncol* 2015;**38**:195–204
24. Xia R, Sheng X, Xu X, Yu C, Lu H. Hesperidin induces apoptosis and G0/G1 arrest in human non-small cell lung cancer A549 cells. *Int J Mol Med* 2018;**41**:464–72
25. Wang Y, Yu H, Zhang J, Gao J, Ge X, Lou G. Hesperidin Inhibits Hela Cell Proliferation through Apoptosis Mediated by Endoplasmic Reticulum Stress Pathways and Cell Cycle Arrest. *BMC Cancer* 2015;**15**:682
26. Pandey P, Sayyed U, Tiwari RK, Siddiqui MH, Pathak N, Bajpai P. Hesperidin induces ROS-mediated apoptosis along with cell cycle arrest at G2/M phase in human gall bladder carcinoma. *Nutr Cancer* 2019;**71**:676–87
27. Naz H, Tarique M, Ahamad S, Alajmi MF, Hussain A, Rehman MT, Luqman S, Hassan MI. Hesperidin-CAMKIV interaction and its impact on cell proliferation and apoptosis in the human hepatic carcinoma and neuroblastoma cells. *J Cell Biochem* 2019;**120**:15119–30
28. Saiprasad G, Chitra P, Manikandan R, Sudhandiran G. Hesperidin induces apoptosis and triggers autophagic markers through inhibition of Aurora-a mediated phosphoinositide-3-kinase/akt/mammalian target of rapamycin and glycogen synthase kinase-3 beta signalling Cascades in experimental Colon carcinogenesis. *Eur J Cancer* 2014;**50**:2489–507
29. Adan A, Baran Y. Fistein and hesperetin induced apoptosis and cell cycle in chronic myeloid leukemia cells accompanied by modulation of cellular signaling. *Tumour Biol* 2015;**37**:5781–95
30. Siddiqi A, Hasan SK, Nafees S, Rashid S, Saidullah B, Sultana S. Chemopreventive efficacy of hesperidin against chemically induced nephrotoxicity and renal carcinogenesis via amelioration of oxidative stress and modulation of multiple molecular pathways. *Exp Mol Pathol* 2015;**99**:641–53
31. Du GY, He SW, Zhang L, Sun CX, Mi LD, Sun ZG. Hesperidin exhibits in vitro and in vivo antitumor effects in human osteosarcoma MG-63 cells and xenograft mice models via inhibition of cell migration and invasion, cell cycle arrest and induction of mitochondrial-mediated apoptosis. *Oncol Lett* 2018;**16**:6299–306
32. Mahmoud AM, Mohammed HM, Khadrawy SM, Galaly SR. Hesperidin protects against chemically induced hepatocarcinogenesis via modulation of Nrf2/ARE/HO-1, PPARgamma and TGF-beta1/Smad3 signaling, and amelioration of oxidative stress and inflammation. *Chem Biol Interact* 2017;**277**:146–58
33. Fernandez-Bedmar Z, Anter J, Alonso-Moraga A, Martin de Las Mulas J, Millan-Ruiz Y, Guil-Luna S. Demethylating and anti-hepatocarcinogenic potential of hesperidin, a natural polyphenol of citrus juices. *Mol Carcinog* 2017;**56**:1653–62
34. Zaghloul RA, Elsherbiny NM, Kenawy HI, El-Karef A, Eissa LA, El-Shishtawy MM. Hepatoprotective effect of hesperidin in hepatocellular carcinoma: involvement of Wnt signaling pathways. *Life Sci* 2017;**185**:114–25
35. Nandakumar N, Jayaprakash R, Balasubramanian MP. Influence of hesperidin on renal cell surface glycoprotein content, nucleic acids, lysosomal enzymes and macromolecules against 7, 12-dimethylbenz [a] anthracene induced experimental breast carcinoma. *J Exp Ther Oncol* 2012;**9**:265–80
36. Huang SM, Tsai SY, Lin JA, Wu CH, Yen GC. Cytoprotective effects of hesperetin and hesperidin against amyloid β -induced impairment of glucose transport through downregulation of neuronal autophagy. *Mol Nutr Food Res* 2012;**56**:601–9
37. Siddiqi A, Saidullah B, Sultana S. Anti-carcinogenic effect of hesperidin against renal cell carcinoma by targeting COX-2/PGE2 pathway in Wistar rats. *Environ Toxicol* 2018;**33**:1069–77
38. He S, Wang X, Zhong Y, Tang L, Zhang Y, Ling Y, Tan Z, Yang P, Chen A. Hesperetin post-treatment prevents rat cardiomyocytes from hypoxia/reoxygenation injury in vitro via activating PI3K/akt signaling pathway. *Biomed Pharmacother* 2017;**91**:1106–12
39. Kamaraj S, Anandakumar P, Jagan S, Ramakrishnan G, Devaki T. Modulatory effect of hesperidin on benzo(a)pyrene induced experimental lung carcinogenesis with reference to COX-2, MMP-2 and MMP-9. *Eur J Pharmacol* 2010;**649**:320–7

40. Hosseinimehr SJ, Jalayer Z, Naghshvar F, Mahmoudzadeh A. Hesperidin inhibits cyclophosphamide-induced tumor growth delay in mice. *Integr Cancer Ther* 2012;**11**:251-6
41. Nandakumar N, Balasubramanian MP. Hesperidin a citrus bioflavonoid modulates hepatic biotransformation enzymes and enhances intrinsic antioxidants in experimental breast cancer rats challenged with 7, 12-dimethylbenz (a) anthracene. *J Exp Ther Oncol* 2012;**9**:321-35
42. Kamaraj S, Ramakrishnan G, Anandakumar P, Jagan S, Devaki T. Antioxidant and anticancer efficacy of hesperidin in benzo(a)pyrene induced lung carcinogenesis in mice. *Invest New Drugs* 2009;**27**:214-22
43. Shanmugam MK, Warriar S, Kumar AP, Sethi G, Arfuso F. Potential role of natural compounds as anti-angiogenic agents in cancer. *Curr Vasc Pharmacol* 2017;**15**:503-19
44. Siveen KS, Ahn KS, Ong TH, Shanmugam MK, Li F, Yap WN, Kumar AP, Fong CW, Terganokar V, Hui KM, Sethi G. Y-tocotrienol inhibits angiogenesis-dependent growth of human hepatocellular carcinoma through abrogation of AKT/mTOR pathway in an orthotopic mouse model. *Oncotarget* 2014;**5**:1897-911
45. Lentini A, Forni C, Provenzano B, Beninati S. Enhancement of transglutaminase activity and polyamine depletion in B16-F10 melanoma cells by flavonoids naringenin and hesperitin correlate to reduction of the in vivo metastatic potential. *Amino Acids* 2007;**32**:95-100
46. Yeh MH, Kao ST, Hung CM, Liu CJ, Lee KH, Yeh CC. Hesperidin inhibited acetaldehyde-induced matrix metalloproteinase-9 gene expression in human hepatocellular carcinoma cells. *Toxicol Lett* 2009;**184**:204-10
47. Lee KH, Yeh MH, Kao ST, Hung CM, Liu CJ, Huang YY, Yeh CC. The inhibitory effect of hesperidin on tumor cell invasiveness occurs via suppression of activator protein 1 and nuclear factor-kappaB in human hepatocellular carcinoma cells. *Toxicol Lett* 2010;**194**:42-9
48. Yang Y, Wolfram J, Shen H, Fang X, Ferrari M. Hesperetin: an inhibitor of the transforming growth factor-beta (TGF-beta) signaling pathway. *Eur J Med Chem* 2012;**58**:390-5
49. Kim GD. Hesperidin inhibits vascular formation by blocking the AKT/mTOR signaling pathways. *Prev Nutr Food Sci* 2015;**20**:221-9
50. Zhao X, Liu J, Feng L, Ge S, Yang S, Chen C, Li X, Peng L, Mu Y, Wang Y, Gu D, Guo Y, Lin G, Deng B, Cheng Z, Cai D. Anti-angiogenic effects of qingdu granule on breast cancer through inhibiting NFAT signaling pathway. *J Ethnopharmacol* 2018;**222**:261-9
51. Lee J, Lee J, Kim M, Kim JH. Fermented extraction of citrus unshiu peel inhibits viability and migration of human pancreatic cancers. *J Med Food* 2018;**21**:5-12
52. Xia R, Xu G, Huang Y, Sheng X, Xu X, Lu H. Hesperidin suppresses the migration and invasion of non-small cell lung cancer cells by inhibiting the SDF-1/CXCR-4 pathway. *Life Sci* 2018;**201**:111-20
53. Wang Y, Liu S, Dong W, Qu X, Huang C, Yan T, Du J. Combination of Hesperetin and Platinum Enhances Anticancer Effect on Lung Adenocarcinoma. *Biomed Pharmacother* 2019;**113**:108779
54. Lee J, Kim D-H, Kim J H. Combined Administration of Naringenin and Hesperetin with Optimal Ratio Maximizes the anti-Cancer Effect in Human Pancreatic Cancer via down Regulation of FAK AND p38 Signaling Pathway. *Phytomedicine* 2019;**58**:152762
55. Banjerdpongchai R, Wudtiwai B, Khaw-On P, Rachakhom W, Duangnil N, Kongtawelert P. Hesperidin from citrus seed induces human hepatocellular carcinoma HepG2 cell apoptosis via both mitochondrial and death receptor pathways. *Tumour Biol* 2016;**37**:227-37
56. Lee KA, Lee SH, Lee YJ, Baeg SM, Shim JH. Hesperidin induces apoptosis by inhibiting Sp1 and its regulatory protein in MSTO-211H cells. *Biomol Ther* 2012;**20**:273-9
57. Zhao J, Li Y, Gao J, De Y. Hesperidin inhibits ovarian cancer cell viability through endoplasmic reticulum stress signaling pathways. *Oncol Lett* 2017;**14**:5569-74
58. Ghorbani A, Nazari M, Jeddi-Tehrani M, Zand H. The citrus flavonoid hesperidin induces p53 and inhibits NF-kappaB activation in order to trigger apoptosis in NALM-6 cells: involvement of PPARgamma-dependent mechanism. *Eur J Nutr* 2012;**51**:39-46
59. Al-Ashaal HA, El-Sheltawy ST. Antioxidant capacity of hesperidin from citrus peel using electron spin resonance and cytotoxic activity against human carcinoma cell lines. *Pharm Biol* 2011;**49**:276-82
60. Barber GN. STING: infection, inflammation and cancer. *Nat Rev Immunol* 2015;**15**:760-70
61. Goldberg JE, Schwertfeger KL. Proinflammatory cytokines in breast cancer: mechanisms of action and potential targets for therapeutics. *Curr Drug Targets* 2010;**11**:1133-46
62. Mantovani A. Molecular pathways linking inflammation and cancer. *Curr Mol Med* 2010;**10**:369-73
63. Chai EZ, Siveen KS, Shanmugam MK, Arfuso F, Sethi G. Analysis of the intricate relationship between chronic inflammation and cancer. *Biochem J* 2015;**468**:1-15
64. Shanmugam MK, Sethi G. Role of epigenetics in inflammation-associated diseases. *Subcell Biochem* 2013;**61**:627-57
65. Sethi G, Shanmugam MK, Ramachandran L, Kumar AP, Tergaonkar V. Multifaceted link between cancer and inflammation. *Biosci Rep* 2012;**32**:1-15
66. Blaser H, Dostert C, Mak TW, Brenner D. TNF and ROS crosstalk in inflammation. *Trends Cell Biol* 2016;**26**:249-61
67. Lawrence T, Gilroy DW, Colville-Nash PR, Willoughby DA. Possible new role for NF-kappaB in the resolution of inflammation. *Nat Med* 2001;**7**:1291-7
68. Minami T, Aird WC. Endothelial cell gene regulation. *Trends Cardiovasc Med* 2005;**15**:174-84
69. Nizamutdinova IT, Jeong JJ, Xu GH, Lee SH, Kang SS, Kim YS, Chang KC, Kim HJ. Hesperidin, hesperidin methyl chalone and phellopterin from poncirus trifoliata (rutaceae) differentially regulate the expression of adhesion molecules in tumor necrosis factor-alpha-stimulated human umbilical vein endothelial cells. *Int Immunopharmacol* 2008;**8**:670-8
70. Zarubin T, Han J. Activation and signaling of the p38 MAP kinase pathway. *Cell Res* 2005;**15**:11-8
71. Zhu Y, Zhu M, Lance P. iNOS signaling interacts with COX-2 pathway in colonic fibroblasts. *Exp Cell Res* 2012;**318**:2116-27
72. Colotta F, Allavena P, Sica A, Garlanda C, Mantovani A. Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis* 2009;**30**:1073-81
73. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* 2010;**140**:883-99
74. Lorzadeh E, Ramezani-Jolfaie N, Mohammadi M, Khoshbakht Y, Salehi-Abargouei A. The effect of hesperidin supplementation on inflammatory markers in human adults: a systematic review and meta-analysis of randomized controlled clinical trials. *Chem Biol Interact* 2019;**307**:8-15
75. Li X, Xie X, Zhang L, Meng Y, Li N, Wang M, Zhai C, Liu Z, Di T, Zhang L, Li P. Hesperidin inhibits keratinocyte proliferation and imiquimod-induced psoriasis-like dermatitis via the IRS-1/ERK1/2 pathway. *Life Sci* 2019;**219**:311-21
76. Parhiz H, Roohbakhsh A, Soltani F, Rezaee R, Iranshahi M. Antioxidant and anti-inflammatory properties of the citrus flavonoids hesperidin and hesperetin: an updated review of their molecular mechanisms and experimental models. *Phytother Res* 2015;**29**:323-31
77. Rice-Evans CA, Miller NJ, Paganga G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radic Biol Med* 1996;**20**:933-56
78. Heim KE, Tagliaferro AR, Bobilya DJ. Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships. *J Nutr Biochem* 2002;**13**:572-84
79. Wilmsen PK, Spada DS, Salvador M. Antioxidant activity of the flavonoid hesperidin in chemical and biological systems. *J Agric Food Chem* 2005;**53**:4757-61
80. Kamaraj S, Anandakumar P, Jagan S, Ramakrishnan G, Devaki T. Hesperidin attenuates mitochondrial dysfunction during benzo(a) pyrene-induced lung carcinogenesis in mice. *Fundam Clin Pharmacol* 2011;**25**:91-8
81. Pradeep K, Ko KC, Choi MH, Kang JA, Chung YJ, Park SH. Protective effect of hesperidin, a citrus flavanoglycone, against gamma-radiation-induced tissue damage in Sprague-Dawley rats. *J Med Food* 2012;**15**:419-27

82. Nguyen T, Nioi P, Pickett CB. The Nrf2-antioxidant response element signaling pathway and its activation by oxidative stress. *J Biol Chem* 2009;**284**:13291-5
83. Chen X, Ding HW, Li HD, Huang HM, Li XF, Yang Y, Zhang YL, Pan XY, Huang C, Meng XM, Li J. Hesperetin derivative-14 alleviates inflammation by activating PPAR- γ in mice with CCl₄-induced acute liver injury and LPS-treated RAW264.7 cells. *Toxicol Lett* 2017;**274**:51-63
84. Kong LN, Lin X, Huang C, Ma TT, Meng XM, Hu CJ, Wang QQ, Liu YH, Shi QP, Li J. Hesperetin derivative-12 (HDND-12) regulates macrophage polarization by modulating JAK2/STAT3 signaling pathway. *Chin J Nat Med* 2019;**17**:122-30
85. Roohbakhsh A, Parhiz H, Soltani F, Rezaee R, Iranshahi M. Neuropharmacological properties and pharmacokinetics of the citrus flavonoids hesperidin and hesperetin - a mini-review. *Life Sci* 2014;**113**:1-6
86. Banjerdpongchai R, Khaw-On P. Terpinen-4-ol induces autophagic and apoptotic cell death in human leukemic HL-60 cells. *Asian Pac J Cancer Prev* 2013;**14**:7537-42
87. Banjerdpongchai R, Punyati P, Nakrob A, Pompimon W, Kongtawelert P. 4'-Hydroxycinnamaldehyde from *Alpinia galanga* (Linn.) induces human leukemic cell apoptosis via mitochondrial and endoplasmic reticulum stress pathways. *Asian Pac J Cancer Prev* 2011;**12**:593-8
88. Lin W, Tongyi S. Role of bax/bcl-2 family members in green tea polyphenol induced necroptosis of p53-deficient Hep3B cells. *Tumour Biol* 2014;**35**:8065-75
89. Korga A, Ostrowska M, Jozefczyk A, Iwan M, Wojcik R, Zgorka G, Herbet M, Vilarrubla G G, Dudka J. Apigenin and Hesperidin Augment the Toxic Effect of Doxorubicin against HepG2 Cells. *BMC Pharmacol Toxicol* 2019;**20**:22
90. Febriansah R, Putri DD, Sarmoko Nurulita NA, Meiyanto E, Nugroho AE. Hesperidin as a preventive resistance agent in MCF-7 breast cancer cells line resistance to doxorubicin. *Asian Pac J Trop Biomed* 2014;**4**:228-33
91. Desai UN, Shah KP, Mirza SH, Panchal DK, Parikh SK, Rawal RM. Enhancement of the cytotoxic effects of cytarabine in synergism with hesperidine and silibinin in acute myeloid leukemia: an in-vitro approach. *J Cancer Res Ther* 2015;**11**:352-7
92. Khamis AAA, Ali EMM, El-Moneim MAA, Abd-Alhaseeb MM, El-Magd MA, Salim EI. Hesperidin, piperine and bee venom synergistically potentiate the anticancer effect of tamoxifen against breast cancer cells. *Biomed Pharmacother* 2018;**105**:1335-43
93. Dokumacioglu E, Iskender H, Sen T M, Ince I, Dokumacioglu A, Kanbay Y, Erbas E, Saral S. The Effects of Hesperidin and Quercetin on Serum Tumor Necrosis Factor-Alpha and Interleukin-6 Levels in Streptozotocin-Induced Diabetes Model. *Pharmacogn Mag* 2018;**14**:167-73
94. Gabizon AA, Patil Y, La-Beck NM. New insights and evolving role of pegylated liposomal doxorubicin in cancer therapy. *Drug Resist Updat* 2016;**29**:90-106
95. Minotti G, Menna P, Salvatorelli E, Cairo G, Gianni L. Anthracyclines: molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. *Pharmacol Rev* 2004;**56**:185-229
96. Agudelo D, Bourassa P, Berube G, Tajmir-Riahi HA. Review on the binding of anticancer drug doxorubicin with DNA and tRNA: structural models and antitumor activity. *J Photochem Photobiol B Biol* 2016;**158**:274-9
97. Lin S, Hoffmann K, Schemmer P. Treatment of hepatocellular carcinoma: a systematic review. *Liver Cancer* 2012;**1**:144-58
98. Leung TW, Johnson PJ. Systemic therapy for hepatocellular carcinoma. *Semin Oncol* 2001;**28**:514-20
99. Iskender H, Dokumacioglu E, Sen T M, Ince I, Kanbay Y, Saral S. The Effect of Hesperidin and Quercetin on Oxidative Stress, NF-Kb and SIRT1 Levels in a STZ-Induced Experimental Diabetes Model. *Biomed Pharmacother* 2017;**90**:500-8
100. Siddiqui IA, Sanna V. Impact of nanotechnology on the delivery of natural products for cancer prevention and therapy. *Mol Nutr Food Res* 2016;**60**:1330-41
101. Wang S, Zhang J, Chen M, Wang Y. Delivering flavonoids into solid tumors using nanotechnologies. *Expert Opin Drug Deliv* 2013;**10**:1411-28
102. Majumdar S, Srirangam R. Solubility, stability, physicochemical characteristics and in vitro ocular tissue permeability of hesperidin: a natural bioflavonoid. *Pharm Res* 2009;**26**:1217-25
103. Duranoglu D, Uzunoglu D, Mansuroglu B, Arasoglu T, Derman S. Synthesis of hesperetin-loaded PLGA nanoparticles by two different experimental design methods and biological evaluation of optimized nanoparticles. *Nanotechnology* 2018;**29**:395603
104. Nie S, Xing Y, Kim GJ, Simons JW. Nanotechnology applications in cancer. *Annu Rev Biomed Eng* 2007;**9**:257-88
105. Merisko-Liversidge E, Liversidge GG, Cooper ER. Nanosizing: a formulation approach for poorly-water-soluble compounds. *Eur J Pharm Sci* 2003;**18**:113-20
106. Gu SF, Wang LY, Tian YJ, Zhou ZX, Tang JB, Liu XR, Jiang HP, Shen YQ. Enhanced water solubility, antioxidant activity, and oral absorption of hesperetin by D-alpha-tocopheryl polyethylene glycol 1000 succinate and phosphatidylcholine. *J Zhejiang Univ Sci B* 2019;**20**:273-81
107. Ansar S, Abudawood M, Alaraj ASA, Hamed SS. Hesperidin alleviates zinc oxide nanoparticle induced hepatotoxicity and oxidative stress. *BMC Pharmacol Toxicol* 2018;**19**:65
108. Ferrari PC, Correia MK, Somer A, Ribeiro MA, Astrath NGC, Sato F, Novatski A. Hesperidin-loaded solid lipid nanoparticles: development and physicochemical properties evaluation. *J Nanosci Nanotechnol* 2019;**19**:4747-57
109. Praveen Kumar P, Sunil Kumar KT, Kavya Nainita M, Sai Tarun A, Raghu Ramdu BG, Deepika K, Pramoda A, Yasmeen C. Cerebroprotective potential of hesperidin nanoparticles against bilateral common carotid artery occlusion reperfusion injury in rats and in silico approaches. *Neurotox Res* 2019; doi: 10.1007/s12640-019-00098-8. [Epub ahead of print]
110. Menezes PD, Frank LA, Lima BD, de Carvalho YM, Serafini MR, Quintans-Junior LJ, Pohlmann AR, Guterres SS, Araujo AA. Hesperetin-loaded lipid-core nanocapsules in polyamide: a new textile formulation for topical drug delivery. *Int J Nanomedicine* 2017;**12**:2069-79
111. Amani H, Ajami M, Maleki SN, Pazoki-Toroudi H, Daglia M, Sokeng AJ, Di Lorenzo A, Nabavi SF, Devi KP, Nabavi SM. Targeting signal transducers and activators of transcription (STAT) in human cancer by dietary polyphenolic antioxidants. *Biochimie* 2017;**142**:63-79
112. Ren DY, Xu T, Li R, Huang C, Huang Y, Li RQ, Li HY, Li J. 5,7,3'-Triacetyl hesperetin suppresses adjuvant-induced arthritis in rats through modulating JAK2/STAT3 pathway. *Am J Chin Med* 2013;**41**:601-14
113. Li R, Cai L, Ren DY, Xie XF, Hu CM. Li Therapeutic effect of 7, 3'-dimethoxy hesperetin on adjuvant arthritis in rats through inhibiting JAK2-STAT3 signal pathway. *J Int Immunopharmacol* 2012;**14**:157-63
114. Li X, Hu X, Wang J, Xu W, Yi C, Ma R, Jiang H. Inhibition of autophagy via activation of PI3K/akt/mTOR pathway contributes to the protection of hesperidin against myocardial ischemia/reperfusion injury. *Int J Mol Med* 2018;**42**:1917-24
115. Shokoohi M, Shoorei H, Khaki A, Khaki AA, Moghimian M, Abtahi-Eivary SH. Hesperidin attenuated apoptotic-related genes in testicle of a male rat model of varicocele. *Andrologia* 2020;**8**:249-58
116. Shoorei H, Banimohammad M, Kebria MM, Afshar M, Taheri MM, Shokoohi M, Farashah MS, Eftekharzadeh M, Akhiani O, Gaspar R, Pazoki-Toroudi H. Hesperidin improves the follicular development in 3D culture of isolated preantral ovarian follicles of mice. *Exp Biol Med* 2019;**244**:352-61